HaloPlex Target Enrichment System

Automation Protocol

For Illumina Sequencing

Protocol

Version F1, July 2015

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In this Guide...

This guide describes an optimized automation protocol for using the HaloPlex target enrichment system to prepare sequencing library samples for Illumina paired-end multiplexed sequencing platforms. Sample processing steps are automated using the Agilent NGS Bravo Option A.

1 Before You Begin

This chapter contains information (such as procedural notes, safety information, required reagents and equipment) that you should read and understand before you start an experiment.

2 Using the Agilent NGS Bravo for HaloPlex Target Enrichment

This chapter contains an orientation to the Agilent NGS Bravo Option A, an overview of the HaloPlex target enrichment protocol, and considerations for designing HaloPlex experiments for automated processing using the Agilent NGS Bravo Option A.

3 Sample Preparation

This chapter describes the steps of the automated HaloPlex workflow to prepare target-enriched sequencing libraries for the Illumina platform.

4 Appendix: Provisional Adaptor-Dimer Removal Protocol

This chapter describes a protocol used to remove adaptor-dimer (125 bp) molecules that may be observed for some designs.

5 Reference

This chapter contains reference information, including component kit contents and index sequences.

What's New in Version F1

- Support for ClearSeq Disease Research Panels (see Table 2 on page 10 and Table 34 on page 85)
- Updates to format of hybridization instructions (see Table 18 on page 49)
- Update to name of indexing primers (see Table 35 on page 86)

What's New in Version F0

- Updated product labeling statement

What's New in Version E0

- Support for kits supplied with either of two indexing primer configurations.

  Kits with revised index configuration (typically received December, 2014 or later) include indexing primers A01 through H12 provided in a blue plate. For kit content details see page 84. For nucleotide sequences of the 8-bp indexes in this revised configuration, see Table 37 on page 88.

  Kits with original index configuration (typically received before December, 2014), include indexing primers 1–96 provided in a clear plate. For kit content details see page 89. For nucleotide sequences of the 8-bp indexes in this original configuration, see Table 41 on page 92 through Table 46 on page 97.

  Protocol steps for indexing using primers provided in either configuration are identical.

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1 Before You Begin

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Make sure you read and understand the information in this chapter and have the necessary equipment and reagents listed before you start an experiment.

Procedural Notes

- The 96 reaction kit contains enough reagents to prepare master mixes for four runs of 3 columns of samples (24 samples) per run. When processing samples using runs with fewer than 24 samples, some reagents may be depleted before 96 samples are run.
- The HaloPlex protocol is optimized for digestion of 200 ng of genomic DNA (split among 8 different restriction digestion reactions) plus 25 ng excess DNA, for a total of 225 ng genomic DNA. Using lower amounts of DNA in the enrichment protocol can adversely affect your results. Use a fluorometry-based DNA quantitation method, such as PicoGreen stain or Qubit fluorometry to quantify the DNA starting material.
- Always keep pre-amplification and post-amplification DNA samples in separate work areas. Perform the enrichment procedure in the pre-amplification area. Open and store the amplified, enriched DNA samples only in the post-amplification area.
- Possible stopping points, where DNA samples may be stored between steps, are marked in the protocol. Store the samples at –20°C, but do not subject the samples to multiple freeze/thaw cycles.
- Ensure that master mixes are thoroughly mixed, by pipetting up-and-down or by gentle vortexing, before distributing to the samples.
- In general, follow Biosafety Level 1 (BL1) safety rules.

Safety Notes

CAUTION

- Wear appropriate personal protective equipment (PPE) when working in the laboratory.

Required Reagents

Table 1 Required Reagents for HaloPlex Target Enrichment

Description	Vendor and part number
HaloPlex Target Enrichment System Kit **or** ClearSeq Target Enrichment System Kit	Select the appropriate kit for your probe design from Table 2
Herculase II Fusion Enzyme with dNTPs (100 mM; 25 mM for each nucleotide), 200 reactions	Agilent p/n 600677
Nuclease-free Water (not DEPC-treated)	Ambion Cat #AM9930
Agencourt AMPure XP Kit 5 mL 60 mL 450 mL	Beckman Coulter Genomics p/n A63880 p/n A63881 p/n A63882
10 M NaOH, molecular biology grade	Sigma, p/n 72068
2 M acetic acid	Sigma, p/n A8976
10 mM Tris-HCl, pH 8.0 or 10 mM Tris-acetate, pH 8.0	General laboratory supplier
100% Ethanol, molecular biology grade	Sigma-Aldrich p/n E7023
Quant-iT dsDNA BR Assay Kit, for use with the Qubit fluorometer 100 assays, 2-1000 ng 500 assays, 2-1000 ng	 Life Technologies p/n Q32850 Life Technologies p/n Q32853

Before ordering a HaloPlex Target Enrichment System Reagent Kit, use Agilent's SureDesign tool at www.agilent.com/genomics/suredesign to design a custom HaloPlex probe or to select a pre-designed ClearSeq disease research probe. Reagent kit ordering information is supplied as part of the SureDesign process and is summarized in Table 2 below.

Table 2 HaloPlex and ClearSeq Target Enrichment System Kits for Illumina Sequencing

	Part Number
Probe Design	**96 Reactions**
HaloPlex Custom Panel Tier 1*, ILMFST	G9901B
HaloPlex Custom Panel Tier 2†, ILM	G9911B
HaloPlex Custom Panel Tier 3‡, ILM	G9921B
ClearSeq Cancer, ILM	G9903B
ClearSeq Cardiomyopathy, ILM	G9908B
ClearSeq AML, ILM	G9913B
ClearSeq ICCG, ILM	G9953B**
ClearSeq Connective Disorder, ILM	G9953B**
ClearSeq Arrhythmia, ILM	G9953B**
ClearSeq Noonan Syndrome, ILM	G9953B**
ClearSeq Chromosome X, ILM	G9953B**

* Tier 1 designs are 1-500 kb and up to 20,000 probes.

† Tier 2 designs are 0.5-2.5 Mb OR 1-500 kb with >20,000 probes.

‡ Tier 3 designs are 2.6 Mb-5 Mb.

** Select the appropriate made-to-order probe option in SureDesign.

NOTE Kits contain enough reagents for 96 reactions total, including one or more control reactions using Enrichment Control DNA (ECD) samples. Each run of up to 96 samples should include one ECD control enrichment reaction.

Required Equipment

Table 3 Required Equipment for HaloPlex Target Enrichment Automated Protocols

Description	Vendor and part number
Agilent NGS Bravo Option A* with VWorks software version 11.3.0.1195	Contact Agilent Automation Solutions for ordering information: Customerservice.automation@agilent.com
PlateLoc Thermal Microplate Sealer with Small Hotplate	Agilent p/n G5402#226
Robotic Pipetting Tips (Sterile, Filtered, 250 µL)	Agilent p/n 19477-022
Thermal Cycler	Agilent SureCycler 8800, p/n G8800A, 96 well plate module, p/n G8810A, and 384 well plate module, p/n G8820A, or equivalent thermal cycler† and accessories
Eppendorf twin.tec full-skirted 96-well PCR plates	Eppendorf p/n 951020401 or 951020619
Eppendorf twin.tec full-skirted 384-well PCR plates*	Eppendorf p/n 951020702
Eppendorf twin.tec half-skirted 96-well PCR plates‡	Eppendorf p/n 951020303
Thermo Scientific Reservoirs	Thermo Scientific p/n 1064156
Nunc DeepWell Plates, sterile, 1.3-mL well volume	Thermo Scientific p/n 260251
Axygen 96 Deep Well Plate, 2.2 mL, Square Well (waste reservoirs)	Axygen p/n P-2ML-SQ-C E & K Scientific p/n EK-2440
Magnetic separator ** 1.5 mL tube-compatible separator or Conical vial-compatible separator	Life Technologies DynaMag-2 magnet, p/n 12321D or equivalent DynaMag-15 magnet, p/n 12301D or equivalent
Benchtop microcentrifuge	VWR p/n 93000-196, or equivalent
Benchtop plate centrifuge	Labnet International MPS1000 Mini Plate Spinner p/n C1000, or equivalent
P10, P20, P200 and P1000 pipettes	Pipetman P10, P20, P200, P1000 or equivalent
NucleoClean Decontamination Wipes	Millipore p/n 3097
Qubit 2.0 Fluorometer	Life Technologies p/n Q32866
Qubit assay tubes	Life Technologies p/n Q32856
Vortex mixer	General laboratory supplier

* Protocols are also compatible with Agilent NGS Workstation Option B. See page 13 for more information.

† Thermal cycler must have a maximum reaction volume specification of at least 100 µL and be compatible with 0.2 mL tubes.

‡ Compatible with Agilent SureCycler 8800.

** Select the appropriate device based on run size. See page 55 to determine magnetic bead volume to be used for your run size.

Optional Validation Reagents and Equipment

Table 4 Reagents and Equipment for Optional Validation Methods

Description	Vendor and part number
2200 TapeStation Platform and Consumables	
2200 TapeStation	Agilent p/n G2964AA or G2965AA
High Sensitivity D1000 ScreenTape	Agilent p/n 5067-5584
High Sensitivity D1000 Reagents	Agilent p/n 5067-5585
2100 Bioanalyzer Platform and Consumables	
2100 Bioanalyzer Laptop Bundle	Agilent p/n G2943CA
2100 Bioanalyzer Electrophoresis Set	Agilent p/n G2947CA
High Sensitivity DNA Kit	Agilent p/n 5067-4626
Gel Electrophoresis Platform and Consumables	
XCell SureLock Mini-cell	Life Technologies p/n EI0001
Novex 6% Polyacrylamide, TBE Pre-cast Gels	Life Technologies p/n EC62655BOX
Novex TBE Running Buffer, 5X	Life Technologies p/n LC6675
Novex High-density TBE Sample Buffer, 5X	Life Technologies p/n LC6678
GelRed Nucleic Acid Stain, 3X in water	Biotium p/n 41001
DNA molecular weight markers	General laboratory supplier
UV-transilluminator	General laboratory supplier

Using the Agilent NGS Workstation Option B for HaloPlex Automation

HaloPlex target enrichment protocols are compatible with the Agilent NGS Workstation Option B. Depending on the configuration of the system purchased, however, additional adapters may be required. Before initiating experiments, see Table 5 below, and verify that the listed adapters are available for your workstation.

Table 5 Adapter checklist for HaloPlex automation using Agilent NGS Workstation Option B

Adapter Description	Quantity Required for HaloPlex Automation	Agilent part number (single adapter)
384-well plate insert	2	G5498B#60
96-well PCR plate insert (red)*	2	G5498B#13

* If your NGS Workstation Option B system is already equipped with one red insert, purchase one additional insert using the ordering information shown.

2 Using the Agilent NGS Bravo for HaloPlex Target Enrichment

About the Agilent NGS Bravo Option A 16
Overview of the HaloPlex Target Enrichment Procedure 25
Experimental Setup Considerations for Automated Runs 27

This chapter contains an orientation to the Agilent NGS Bravo (Option A), an overview of the HaloPlex target enrichment protocol, and considerations for designing HaloPlex experiments for automated processing using the Agilent NGS Bravo.

About the Agilent NGS Bravo Option A

About the Bravo Platform

The Bravo platform is a versatile liquid handler with a nine plate-location platform deck, suitable for handling 96-well, 384-well, and 1536-well plates. The Bravo platform is controlled by the VWorks Automation Control software. Fitted with a choice of seven interchangeable fixed-tip or disposable-tip pipette heads, it accurately dispenses fluids from 0.1 µL to 250 µL.

CAUTION Before you begin, make sure that you have read and understand operating, maintenance and safety instructions for using your Bravo platform. Refer to the *Bravo Platform User Guide* (G5409-90006) and the *VWorks Software User Guide* (G5415-90063).

Bravo Platform Deck

The protocols in the following sections include instructions for placing plates and reagent reservoirs on specific Bravo deck locations. Use Figure 1 to familiarize yourself with the location numbering convention on the Bravo platform deck.

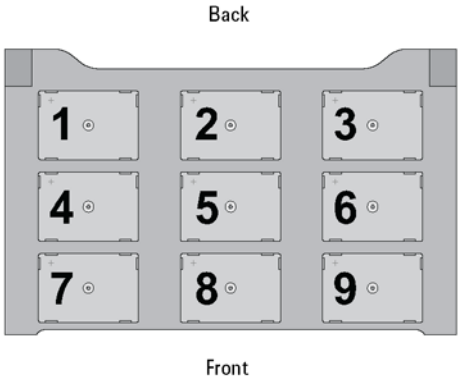


Figure 1 Bravo platform deck

Setting the Temperature of Bravo Deck Heat Blocks

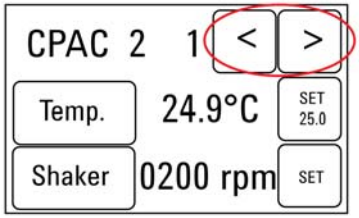
Bravo deck positions 4 and 6 are equipped with Inheco heat blocks, used to incubate sample plates at defined temperatures during the run. Runs that include low-temperature (4°C) or high-temperature (54°C) incubation steps may be expedited by pre-setting the temperature of the affected block before starting the run.

Bravo deck heat block temperatures may be changed using the Inheco Multi TEC Control device touchscreen as described in the steps below. See Table 6 for designations of the heat block-containing Bravo deck positions on the Multi TEC control device.

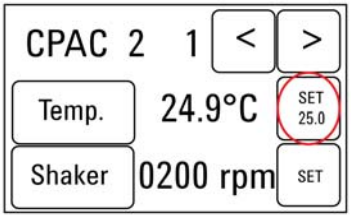
Table 6 Inheco Multi TEC Control touchscreen designations

Bravo Deck Position	Designation on Inheco Multi TEC Control Screen
4	CPAC 2 1
6	CPAC 2 2

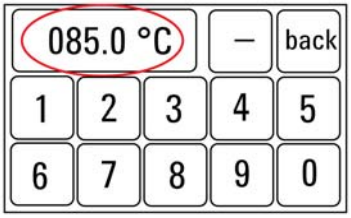
1 Using the arrow buttons, select the appropriate block (CPAC 2 block 1 or CPAC 2 block 2).



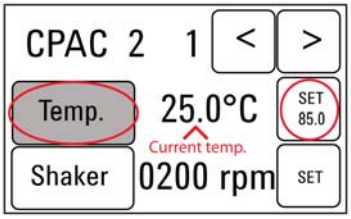
2 To set the temperature of the selected block, press the SET button.



3 Using the numeral pad, enter the desired temperature. The entered temperature appears in the top, left rectangle. Once the correct temperature is displayed, press the rectangle to enter the temperature.



4 Press the Temp button until the new temperature is displayed on the SET button and until the Temp button is darkened, indicating that the selected heat block is heating or cooling to the new temperature setting. The current temperature of the block is indicated in the center of the display.



VWorks Automation Control Software

VWorks software, included with your Agilent NGS Bravo, allows you to control the robot and integrated devices using a PC. The Agilent NGS Bravo Option A is preloaded with VWorks software containing all of the necessary HaloPlex system liquid handling protocols. General instructions for starting up the VWorks software and the included protocols is provided below. Each time a specific VWorks protocol is used in the HaloPlex procedure, any settings required for that protocol are included in the relevant section of this manual.

NOTE

The instructions in this manual are compatible with VWorks software version 11.3.0.1195.

If you have questions about VWorks version compatibility, please contact service.automation@agilent.com.

Logging in to the VWorks software

1 Double-click the VWorks icon or the HaloPlex.VWForm shortcut on the Windows desktop to start the VWorks software.

2 If User Authentication dialog is not visible, click **Log in** on the VWorks window toolbar.

3 In the User Authentication dialog, type your VWorks user name and password, and click **OK**. (If no user account is set up, contact the administrator.)

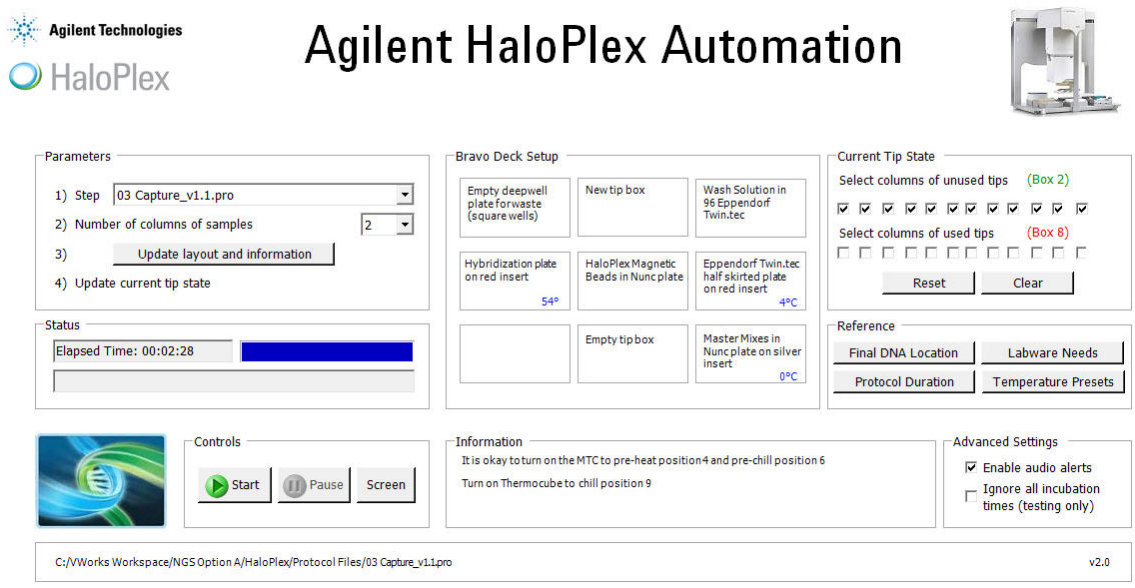
Using the HaloPlex.VWForm to setup and start a run

Use the VWorks form HaloPlex.VWForm, shown below, to set up and start each HaloPlex automation protocol.

1. Open the form using the HaloPlex.VWForm shortcut on your desktop.
2. Use the drop-down menus on the form to select the appropriate HaloPlex workflow step and number of columns of samples for the run.
3. Once all run parameters have been specified on the form, click **Update layout and information.**

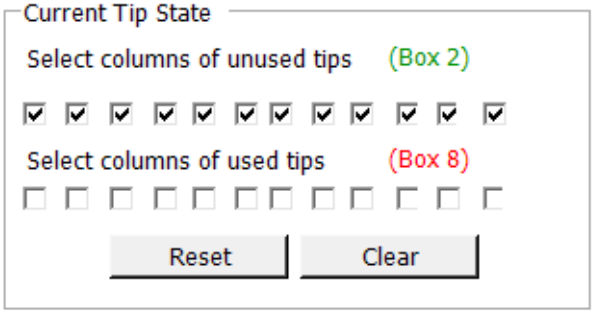
NOTE The displayed protocol will not run unless the **Update layout and information** button has been clicked.

4. The **Bravo Deck Setup** region of the form will then display the required placement of reaction components and labware on the NGS Bravo deck for the specified run parameters.



5 Verify that the **Current Tip State** indicator on the form (shown below) matches the configuration of unused tips in the tip box at Bravo Deck position 2.

For a fresh tip box, containing 12 columns of tips, all positions of the **Current Tip State** unused tip indicator (top portion, Box 2) should be selected, as shown below. Clicking **Reset** selects all columns for position 2.



Also verify that the used tip indicator (bottom portion, Box 8) matches the configuration of used tips in the tip box at Bravo Deck position 8.

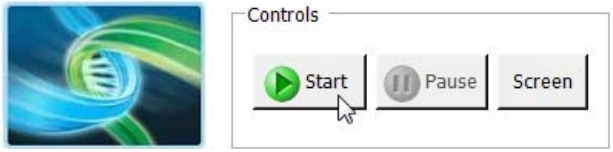
For an empty tip box, all positions of the **Current Tip State** used tip indicator (bottom portion, Box 8) should be cleared, as shown above. Clicking **Reset** clears all columns for position 8.

NOTE

It is important that the **Current Tip State** indicator matches the configuration of tips present at Bravo Deck positions 2 and 8 when initiating the run. Tips that are inappropriately loaded onto the Bravo platform pipette head, or tips missing from the pipette head, will interfere with automated processing steps.

You can use partial tip boxes for HaloPlex automation protocols, as long as positions of available tips are accurately indicated during run setup.

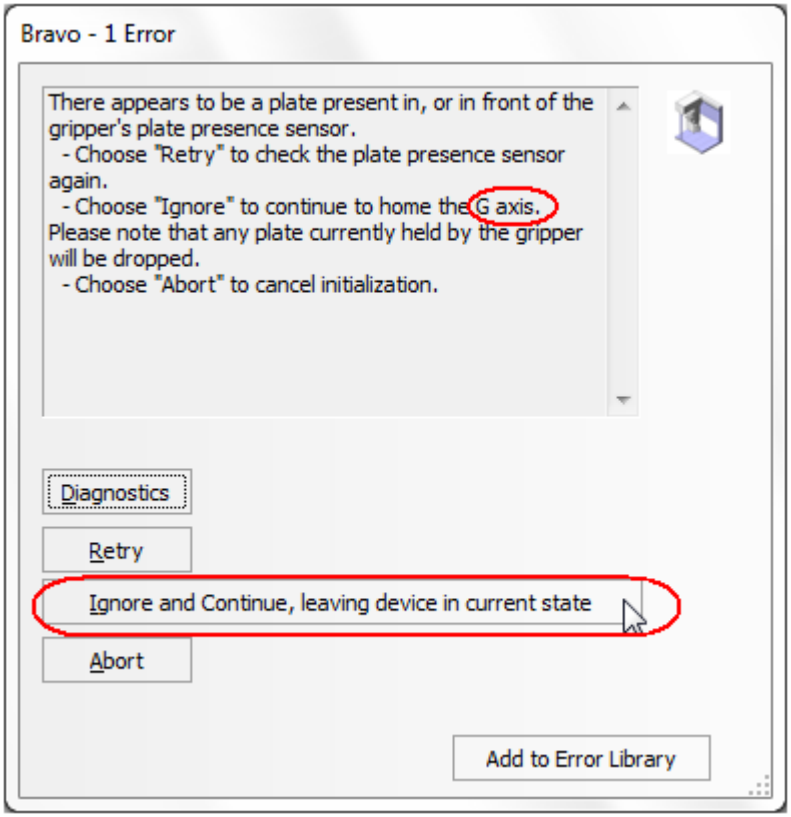
6 After verifying that the NGS Bravo has been set up correctly, click **Start** in the **Controls** section of the form to begin the run. Do not use the Start button on the VWorks Control Toolbar; runs must be initiated using the start button on the HaloPlex.VW Form, shown below.



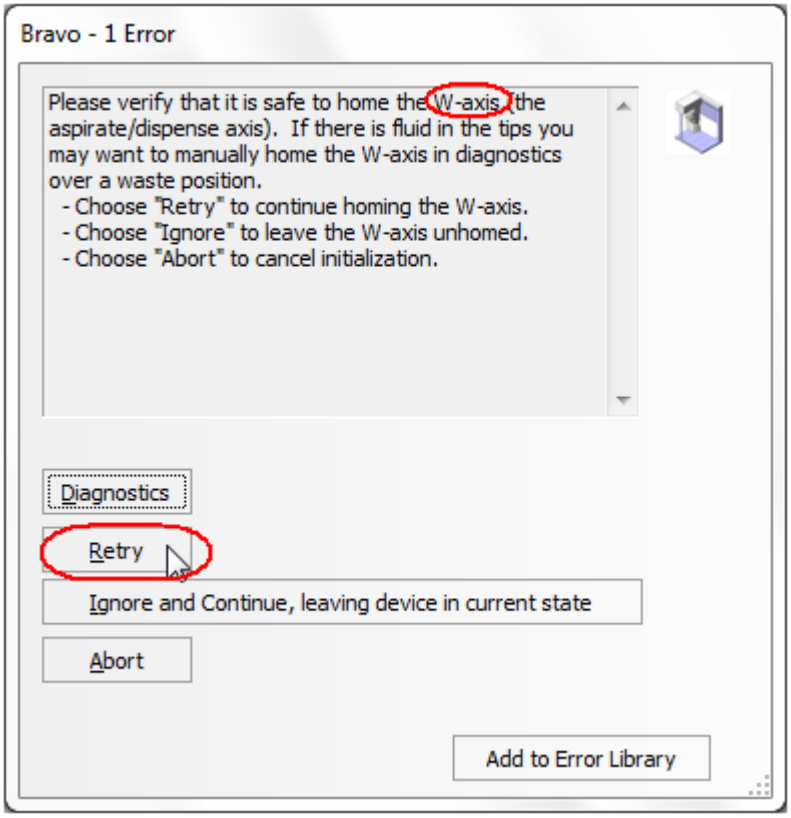
Error messages encountered at start of run

After starting the run, you may see the error messages displayed below. When encountered, make the indicated selections and proceed with the run. Encountering either or both of these error messages is not indicative of a problem with the NGS Bravo or your run setup.

1 If you encounter the G-axis error message shown below, select **Ignore and Continue, leaving device in current state**.


Bravo - 1 Error
There appears to be a plate present in, or in front of the gripper's plate presence sensor.
- Choose "Retry" to check the plate presence sensor again.
- Choose "Ignore" to continue to home the G axis.
Please note that any plate currently held by the gripper will be dropped.
- Choose "Abort" to cancel initialization.
Diagnostics
Retry
Ignore and Continue, leaving device in current state
Abort
Add to Error Library

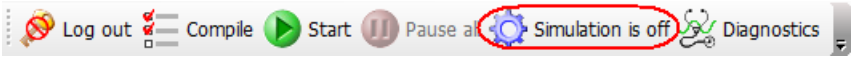
2 If you encounter the W-axis error message shown below, select **Retry.**


Bravo - 1 Error
Please verify that it is safe to home the W-axis (the aspirate/dispense axis). If there is fluid in the tips you may want to manually home the W-axis in diagnostics over a waste position.
- Choose "Retry" to continue homing the W-axis.
- Choose "Ignore" to leave the W-axis unhomed.
- Choose "Abort" to cancel initialization.
Diagnostics
Retry
Ignore and Continue, leaving device in current state
Abort
Add to Error Library

Verifying the Simulation setting

VWorks software may be run in simulation mode, during which commands entered on screen are not completed by the NGS Bravo. If NGS Bravo devices do not respond when you start a run, verify the simulation mode status in VWorks using the following steps.

1 Verify that **Simulation is off** is displayed on the status indicator (accessible by clicking **View** > **Control Toolbar**).



2 If the indicator displays **Simulation is on,** click the status indicator button to turn off the simulation mode.

NOTE If you cannot see the toolbar above the HaloPlex. VWorks form, click **Screen** in the **Controls** section of the form to exit full screen mode. If the toolbar is still not visible, right-click on the form and then select **Control Toolbar** from the menu.

Overview of the HaloPlex Target Enrichment Procedure

Figure 2 summarizes the HaloPlex target enrichment workflow. For each sample to be sequenced, individual HaloPlex-enriched, indexed libraries are prepared. Depending on the specific sequencing platform used, up to 96 samples can be pooled and sequenced in a single lane.

Table 7 summarizes how the VWorks automation protocols are integrated into the HaloPlex workflow. See the Sample Preparation chapter for complete instructions for use of the VWorks protocols for sample processing.

Table 7 Overview of VWorks protocols used during the workflow

Workflow Step	VWorks Protocol used for Automation
Digest genomic DNA	Digestion.pro
Hybridize to HaloPlex or ClearSeq probe and index samples	Hybridization.pro
Capture and amplify enriched DNA	Capture_v1.1.pro
Purify amplified libraries	Purification_v1.1.pro

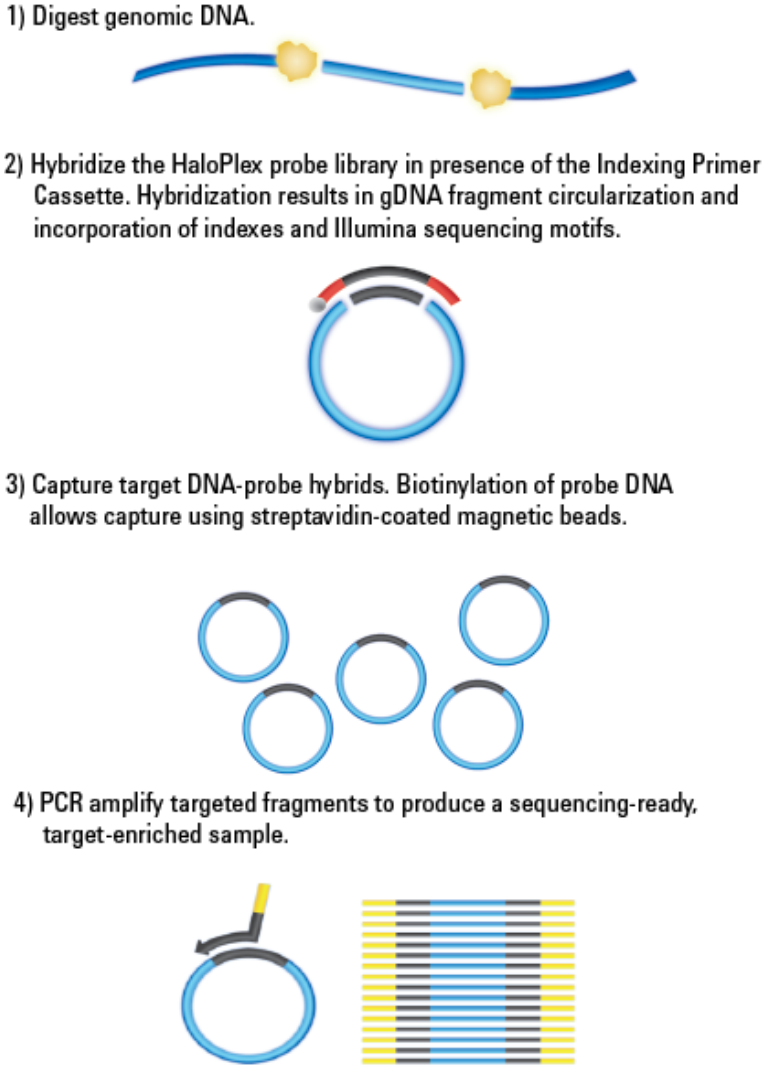

1) Digest genomic DNA.
2) Hybridize the HaloPlex probe library in presence of the Indexing Primer Cassette. Hybridization results in gDNA fragment circularization and incorporation of indexes and Illumina sequencing motifs.
3) Capture target DNA-probe hybrids. Biotinylation of probe DNA allows capture using streptavidin-coated magnetic beads.
4) PCR amplify targeted fragments to produce a sequencing-ready, target-enriched sample.

Figure 2 Overall HaloPlex target-enriched sequencing sample preparation workflow.

Experimental Setup Considerations for Automated Runs

HaloPlex Automated Target Enrichment System runs may include 1, 2, 3, 4, 6, or 12 columns (equivalent to 8, 16, 24, 32, 48, or 96 wells) of gDNA samples to be enriched for sequencing on the Illumina platform. Plan your experiments using complete columns of samples.

Table 8 Columns to Samples Equivalency

Number of Columns Processed	Total Number of Samples Processed
1	8
2	16
3	24
4	32
6	48
12	96

The number of columns or samples that may be processed using the supplied reagents (see page 84) will depend on the experimental design. For greatest efficiency of reagent use, plan experiments using at least 3 columns per run. Each 96-reaction kit contains sufficient reagents for 96 reactions configured as 4 runs of 3 columns of samples per run.

Considerations for Placement of gDNA Samples in 96-well Plates for Automated Processing

- The Agilent NGS Bravo processes samples column-wise beginning at column 1. gDNA samples should be loaded into 96-well plates column-wise, in well order A1 to H1, then A2 to H2, ending with A12 to H12. When processing partial runs with <12 sample columns, do not leave empty columns between sample columns; always load the plate using the left-most column that is available.
- For sample indexing during hybridization to the HaloPlex or ClearSeq probe (see Figure 2), you will need to prepare a separate plate containing the HaloPlex Indexing Primers. Assign the wells to be indexed with their respective indexing primers during experimental design. See the Reference chapter for nucleotide sequences of the 96 indexes used in the HaloPlex Target Enrichment System.

CAUTION

This guide includes information for kits containing two different sets of indexing primers. **Verify that you are referencing the information appropriate for your kit version before you proceed.**

Kits with indexing primers supplied in a blue plate include 8-bp indexes A01 through H12. See page 87 through page 88 for indexing primer A01–H12 plate map and nucleotide sequence information.

Kits with indexing primers supplied in a clear plate include 8-bp indexes 1 through 96. See page 91 through page 97 for indexing primer 1–96 plate map and nucleotide sequence information.

Protocol steps for indexing using primers provided in either configuration are identical.

Considerations for Equipment Setup

- Some workflow steps require the rapid transfer of sample plates between the Bravo deck and a thermal cycler. Locate your thermal cycler in close proximity to the Agilent NGS Bravo to allow rapid and efficient plate transfer.

- Some workflow steps require that the sample plate be sealed, then centrifuged to collect any dispersed liquid, before being transfered between instruments. To maximize efficiency, locate the PlateLoc thermal microplate sealer and the centrifuge in close proximity to the NGS Bravo and thermal cycler.

Run Time Considerations

Before you begin, refer to the Certificate of Analysis provided with Box 1 of your kit to determine the hybridization duration appropriate for your design. After reviewing the duration of this and other steps in the protocol, plan the start time for your experiment accordingly.

Designs containing <20,000 probes, including the disease research panels (Cancer Research, Cardiomyopathy Research, and ClearSeq AML) use a 3-hour hybridization time. For these designs DNA digestion through PCR protocols (see Figure 2) are typically run on the same day with the DNA digestion protocol initiated early in the day.

Designs containing >20,000 probes use a 16-hour hybridization time, which is typically completed overnight. Calculate the appropriate start time for the DNA digestion protocol, based on your run size and the run time estimates provided in the HaloPlex form in the VWorks software (HaloPlex.VWForm), to allow overnight hybridization.

3 Sample Preparation

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Step 2. Hybridize digested DNA to HaloPlex or ClearSeq probe for target enrichment and sample indexing 44
Step 3. Capture and amplify the target DNA 50
Step 4. Purify the amplified target DNA 65
Step 5. Validate enrichment and quantify enriched target DNA 69
Step 6. Pool samples with different indexes for multiplexed sequencing 73

This section contains instructions for gDNA library target enrichment for sequence analysis using the Illumina platform. For each sample to be sequenced, an individual target-enriched, indexed library is prepared.

The target region can vary from 1 kb to 5 Mb. Custom HaloPlex probes must be designed before purchasing the kit using Agilent's SureDesign tool at www.agilent.com/genomics/suredesign.

The HaloPlex Target Enrichment System amplifies thousands of targets in the same reaction, incorporating standard Illumina paired-end sequencing motifs in the process. During hybridization, each sample can be uniquely indexed, allowing for pooling of up to 96 samples per sequencing lane.

Step 1. Digest genomic DNA with restriction enzymes

In this step, gDNA samples are digested by 16 different restriction enzymes to create a library of gDNA restriction fragments. The gDNA is digested in eight different restriction reactions, each containing two restriction enzymes. **The 16 restriction enzymes are provided in two 8-well strip tubes that are distinguished by red and green color markers.** Enzymes are combined from corresponding wells of the red- and green-marked strip tubes to make eight different RE Master Mixes, which are then combined with each DNA sample in the run.

NOTE

Successful enrichment using the protocol in this guide requires high-quality DNA samples. Before you begin, verify that the genomic DNA samples have an OD 260/280 ratio ranging from 1.8 to 2.0. Verify the size distribution of DNA in each DNA preparation by gel electrophoresis. Any smearing below 2.5 kb indicates sample degradation.

For HaloPlex target enrichment of FFPE-derived DNA samples, see Agilent publication no. G9900-90050, available at http://www.genomics.agilent.com. This publication provides a PCR-based protocol for assessment of DNA integrity and provides HaloPlex protocol modifications for improved performance from lower-quality DNA samples.

Prepare the NGS Bravo

1. Gently wipe down the Bravo deck with a NucleoClean decontamination wipe.
2. Place red aluminum inserts on Bravo deck positions 4 and 9.
3. Turn on the chiller, set to 0°C, at position 9 of the Bravo deck. Be sure that the chiller reservoir contains at least 300 mL of 25% ethanol.
4. Place a 384-well adapter insert on Bravo deck position 6.
5. Pre-set the temperature of Bravo deck positions 4 and 6 to 4°C using the Inheco Multi TEC control touchscreen, as described in Setting the Temperature of Bravo Deck Heat Blocks.

NOTE

To expedite thermal cycler warm-up for the restriction digest incubation on page 38, you can enter and initiate the digestion program on the thermal cycler now, and then pause the program until you are instructed to transfer the reaction plate. Be sure to pause the thermal cycler before the initiation of the 37°C incubation segment. Release the pause immediately after transferring the plate to the thermal cycler in step 14 on page 38. Be sure that the 384-well block is in the thermal cycler before initiating the program for warm-up.

Prepare the DNA Sample Source Plate

NOTE

In the protocol below, 200 ng genomic DNA is split among eight different restriction digests, with an additional 25 ng excess DNA included to allow for pipetting losses. Using <225 ng DNA in the enrichment protocol can result in low yield and can potentiate rare allele dropouts. Use a fluorometry-based DNA quantitation method, such as Qubit fluorometry or PicoGreen staining, to accurately quantify the DNA starting material.

1 Use the Qubit dsDNA BR Assay or PicoGreen staining kit to determine the concentration of your gDNA samples.

Follow the manufacturers instructions for the kits and instruments.

2 Prepare the DNA sample plate for the run, containing up to 95 gDNA samples and the Enrichment Control DNA sample, using a full-skirted 96-well Eppendorf twin.tec plate.

NOTE

HaloPlex Automated Target Enrichment System runs may include 1, 2, 3, 4, 6, or 12 columns of the plate. Use full columns of DNA samples for each run.

a In well A1 of a 96-well twin.tec plate, dispense 45 µL of the supplied Enrichment Control DNA (ECD). Store on ice.

b In separate wells of the same 96-well twin.tec plate, dilute 225 ng of each gDNA sample in 45 µL nuclease-free water, for a final DNA concentration of 5 ng/µL. Continue to store on ice.

For automated processing, fill plate wells column-wise in well order A1 to H1, then A2 to H2, ending with A12 to H12.

Prepare the RE Master Mix Source Plate

1 Prepare the appropriate amount of RE Buffer +BSA mixture, according to the table below.

Table 9 Preparation of RE Buffer + BSA mixture for Digestion.pro protocol

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
RE Buffer	34 µL	408 µL	680 µL	952 µL	1224 µL	1768 µL	3536 µL
BSA Solution	0.85 µL	10.2 µL	17 µL	23.8 µL	30.6 µL	44.2 µL	88.4 µL
Total Volume	**34.85 µL**	**418.2 µL**	**697 µL**	**975.8 µL**	**1254.6 µL**	**1812.2 µL**	**3624.4 µL**

2 Obtain the two provided green- and red-marked Enzyme Strips from Box 1. For each strip, label the color-marked tube with A, then continue labeling the remaining tubes with B through H, in order. Keep the strips on ice.

CAUTION It is important to use the restriction enzyme tube strips in the proper orientation when preparing the RE Master Mixes as described below. The red or green color marker on the tube strip and cap strip are used to mark well A of each enzyme strip.

3 In eight individual tubes, prepare the eight Restriction Enzyme Master Mixes A, B, C, D, E, F, G, and H according to the table below. To prepare Master Mix A, combine RE Buffer + BSA from step 1 with the indicated volumes of enzyme solution from well A of the Green Enzyme Strip and from well A of the Red Enzyme Strip. Prepare Master Mixes B–H by repeating this process using enzyme solutions from the corresponding wells B-H of each provided Enzyme Strip.

Table 10 Preparation of RE Master Mixes A–H for Digestion.pro protocol

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
RE Buffer + BSA	4.0	51.0 µL	85.0 µL	119.0 µL	153.0 µL	221.0 µL	442.0 µL
Green Enzyme Strip enzyme A–H	0.5	6.4 µL	10.6 µL	14.9 µL	19.1 µL	27.6 µL	55.25 µL
Red Enzyme Strip enzyme A–H	0.5	6.4 µL	10.6 µL	14.9 µL	19.1 µL	27.6 µL	55.25 µL
Total Volume for each Master Mix A, B, C, D, E, F, G, or H	**5 µL**	**63.8 µL**	**106.2 µL**	**148.8 µL**	**191.2 µL**	**276.2 µL**	**552.5 µL**

NOTE For 1-4 column runs, RE master mixes A-H may be prepared in a 8 x 0.2-mL well strip tube, using a multichannel pipette to transfer volumes from Enzyme Strips 1 and 2 to the RE master mix strip. For 6- or 12-column runs, prepare the master mixes in 1.5-mL tubes.

4 Mix by gentle vortexing and then spin briefly. Keep on ice.

Prepare the RE master mix source plate

1 Aliquot the Restriction Enzyme Master Mixes to a full-skirted 96-well Eppendorf twin.tec plate as shown in Figure 3. Add the volumes indicated in Table 11 of each master mix A–H to each well of the indicated column of the twin.tec plate. Keep the master mixes on ice during the aliquoting steps.

Table 11 Preparation of the RE Master Mix Source Plate for Digestion.pro

Master Mix Solution	Position on Source Plate	Volume of Master Mix added per Well of Source Plate					
		1-Column Runs	2-Column Runs	3-Column Runs	4-Column Runs	6-Column Runs	12-Column Runs
RE Master Mix A	Column 1 (A1-H1)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix B	Column 2 (A2-H2)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix C	Column 3 (A3-H3)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix D	Column 4 (A4-H4)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix E	Column 5 (A5-H5)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix F	Column 6 (A6-H6)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix G	Column 7 (A7-H7)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL
RE Master Mix H	Column 8 (A8-H8)	7.3 µL	12.7 µL	18.0 µL	23.3 µL	33.9 µL	68.4 µL

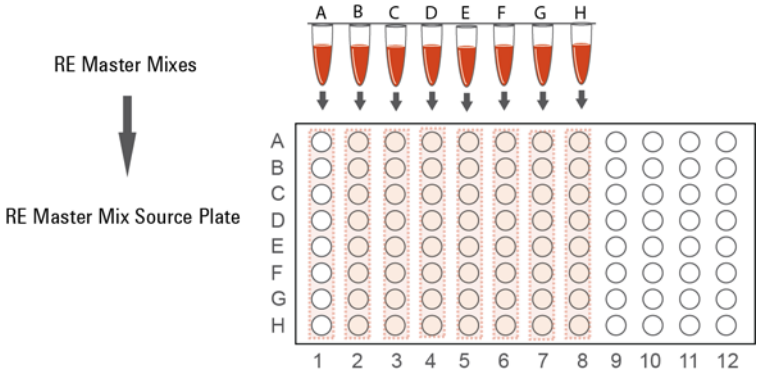


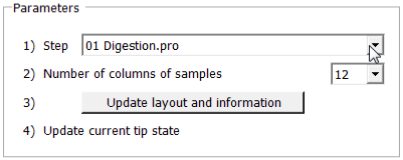
Figure 3 Preparation of the RE Master Mix source plate for automation protocol Digestion.pro.

Load the NGS Bravo and Run the Digestion.pro VWorks Protocol

1 Open the HaloPlex setup form using the HaloPlex.VWForm shortcut on your desktop.

2 Log in to the VWorks software.

3 On the setup form, under **Step**, select **01 Digestion.pro.**



4 Select the number of columns of samples to be processed. Runs must include 1, 2, 3, 4, 6, or 12 columns.

5 Click **Update layout and information.**

6 Load the Bravo deck according to Table 12.

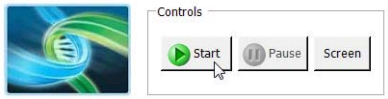
Table 12 Initial Bravo deck configuration for Digestion.pro

Location	Content
1	—(empty)—
2	New tip box
3	For 12-column runs only: Empty 384-well Eppendorf twin.tec plate (no 384-well insert required) For 1- to 6-column runs: empty
4	gDNA samples in full-skirted 96-well Eppendorf twin.tec plate seated on red insert
5	—(empty)—
6	Empty 384-well Eppendorf twin.tec plate seated on 384-well insert
7	—(empty)—
8	Empty tip box
9	RE Master Mix source plate (full-skirted 96-well Eppendorf twin.tec plate) seated on red insert

7 Verify that the NGS Bravo has been set up as displayed in the **Bravo Deck Setup** and **Information** regions of the form.

8 Verify that the **Current Tip State** indicator on the form matches the configuration of unused and used tips in the tip boxes at Bravo Deck positions 2 and 8, respectively. See page 21 for more information on using this segment of the form during the run.

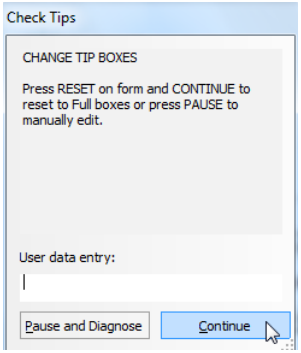
9 When verification is complete, click **Start** to start the run.



NOTE If Bravo devices do not respond when you start the run, but activity is recorded in the Log, verify that VWorks is not running in Simulation mode. See page 24 for more information.

10 When prompted by VWorks as shown below, replace the tip box at position 2 with a new tip box and replace the used tip box at position 8 with an empty tip box. After both tip boxes are in place, click **Reset** under **Current Tip State** on the form. Verify that the tip state was updated and then click **Continue** on the prompt shown below.

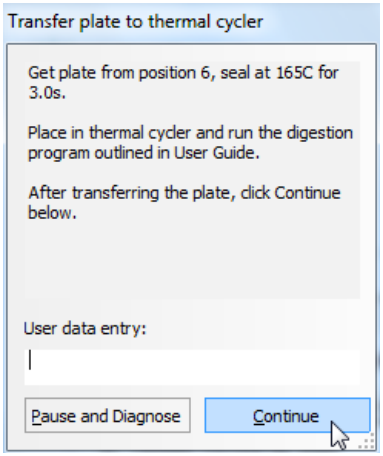
Depending on the run size, you may be prompted to change tip boxes multiple times during the run.


Check Tips
CHANGE TIP BOXES
Press RESET on form and CONTINUE to reset to Full boxes or press PAUSE to manually edit.
User data entry:
Pause and Diagnose
Continue

The NGS Bravo combines each gDNA sample with each RE Master Mix in wells of a 384-well reaction plate. For 1- to 6-column runs, a single 384-well restriction digest plate is prepared; for 12 column runs, two 384-well restriction digest plates are prepared.

11 When the NGS Bravo has finished preparing each 384-well restriction digest plate for the run, you will be prompted by VWorks as shown below.

The final Bravo deck position of the prepared restriction digest plate varies for different run sizes.


Transfer plate to thermal cycler
Get plate from position 6, seal at 165C for 3.0s.
Place in thermal cycler and run the digestion program outlined in User Guide.
After transferring the plate, click Continue below.
User data entry:
Pause and Diagnose
Continue

12 Remove the 384-well plate from the Bravo deck position indicated in the prompt.

13 Seal the sample plate using the PlateLoc Thermal Microplate Sealer, with sealing settings of 165°C and 3.0 sec. Spin the plate briefly to release any bubbles trapped in the liquid.

14 Transfer the sealed plate to a thermal cycler and run the digestion program shown in Table 13, using a heated lid. After transferring the plate, click **Continue** on the prompt.

Table 13 Thermal cycler program for restriction digestion

Step	Temperature	Time
Step 1	37°C	30 minutes
Step 2	8°C	Hold

Preparation of each restriction digest reaction plate takes approximately 30-45 minutes.

For 12 column-runs, the two 384-well plates are prepared sequentially, for a total run time of approximately 90 minutes. Run the thermal cycler digestion program for each plate as soon as prompted. During the 30-minute incubation of plate 1 in the thermal cycler, the NGS Bravo begins preparation of the digestion reactions in plate 2. Once the thermal cycler program is complete for plate 1, store the digested DNA in plate 1 on ice until the Digestion.pro protocol and thermal cycler program for plate 2 is finished.

15 Validate the restriction digestion reaction by electrophoretic analysis of the Enrichment Control DNA (ECD) reactions.

a Transfer 4 µL of each ECD digestion reaction from the wells of the 384-well reaction plate indicated in Table 14 to fresh 0.2-mL PCR tubes. Note that for 12-column runs, four of the eight ECD digests are found on the first 384-well plate, and the remaining four digests are on the second 384-well plate.

Table 14 Position of ECD digestion reactions for obtaining validation samples

Restriction Enzyme Master Mix to be Validated	Position of ECD Digestion Reaction in 384-Well Plates	
	1-6 Column Runs	12-Column Runs (two 384-well plates produced)
RE Master Mix A	A1	A1 (plate 1)
RE Master Mix B	A2	A2 (plate 1)
RE Master Mix C	B1	B1 (plate 1)
RE Master Mix D	B2	B2 (plate 1)
RE Master Mix E	A13	A1 (plate 2)
RE Master Mix F	A14	A2 (plate 2)
RE Master Mix G	B13	B1 (plate 2)
RE Master Mix H	B14	B2 (plate 2)

b Incubate the removed 4-µL samples at 80°C for 5 minutes to inactivate the restriction enzymes.

c Analyze the prepared samples using microfluidic electrophoresis on the 2100 Bioanalyzer (see page 41) or on the 2200 TapeStation (see page 42) or using gel electrophoresis (see page 43).

The ECD sample contains genomic DNA mixed with an 800-bp PCR product that contains restriction sites for all the enzymes used in the digestion protocol. When analyzing validation results, the undigested control should have gDNA bands at >2.5 kbp and a PCR product band at 800 bp. Each of the eight digested ECD samples should have a smear of gDNA restriction fragments between 100 and 2500 bp, overlaid with three predominant bands at approximately 125, 225 and 450 bp. These three bands correspond to the 800-bp PCR product-derived restriction fragments, and precise sizes will differ after digestion in each of the eight RE master mixes.

NOTE

In addition to the three predominant bands at approximately 125, 225 and 450 bp, you may detect additional, minor bands in the digested ECD sample lanes.

Successful digestion is indicated by the appearance of the three predominant bands. The presence of additional minor bands, with relative abundance similar to the additional bands visible in Figure 4, Figure 5 and Figure 6, does not impact enrichment results.

It is acceptable for band intensities in digestion reaction B to be slighltly reduced, compared to the other digestion reactions.

Option 1: Validation by 2100 Bioanalyzer analysis

Use a High Sensitivity DNA Kit (p/n 5067-4626) and the 2100 Bioanalyzer system with 2100 Expert Software (version B.02.07 or higher required to run the High Sensitivity Kit). See the reagent kit guide for general Bioanalyzer system setup instructions.

- Prepare an undigested DNA gel control by combining 0.5 µL of the Enrichment Control DNA stock solution and 3.5 µL of nuclease-free water.
- Prepare the chip, samples and ladder as instructed in the reagent kit guide, using 1 µL of each ECD sample and undigested DNA control for the analysis.
- When loading samples on the chip, load the DNA ladder in the ladder sample well marked on the chip. Load the eight ECD digest samples (A to H) in sample wells 1 to 8, and load the undigested ECD sample in sample well 9. Do not run the undigested ECD control in sample well 1.
- Place the prepared chip into the 2100 Bioanalyzer instrument and start the run within five minutes after preparation.

See Figure 4 for sample Bioanalyzer electrophoresis results.

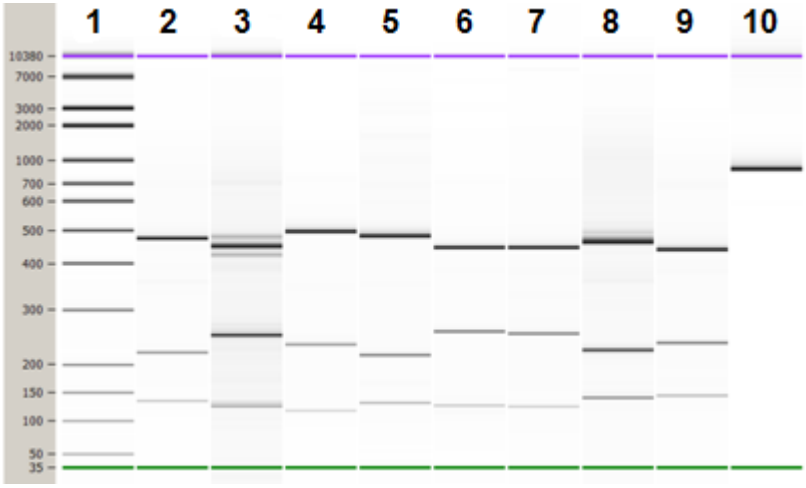


Figure 4 Validation of restriction digestion by 2100 Bioanalyzer system analysis. Lane 1: 50-bp DNA ladder, Lanes 2-9: ECD digestion reactions A–H, Lane 10: Undigested Enrichment Control DNA.

Option 2: Validation by 2200 TapeStation analysis

Use a High Sensitivity D1000 ScreenTape and reagent kit. For more information to do this step, see the *Agilent 2200 TapeStation User Manual* at www.genomics.agilent.com.

- Prepare an undigested DNA gel control by combining 1 µL of the Enrichment Control DNA stock solution and 1 µL of nuclease-free water.
- Prepare the TapeStation samples as instructed in the *2200 TapeStation User Manual*. Use 2 µL of each ECD sample diluted with 2 µL of High Sensitivity D1000 sample buffer in separate wells of a tube strip for the analysis.
- Load the sample tube strip, the High Sensitivity D1000 ScreenTape, and loading tips into the 2200 TapeStation as instructed in the *2200 TapeStation User Manual*. Start the run.

See Figure 5 for sample TapeStation electrophoresis results.

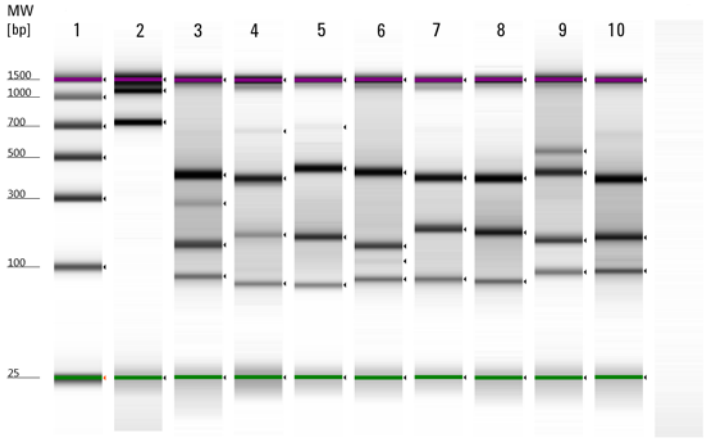


Figure 5 Validation of restriction digestion by 2200 TapeStation analysis. Lane 1: High-Sensitivity Ladder, Lane 2: Undigested Enrichment Control DNA, Lanes 3–10: ECD digestion reactions A–H.

Option 3: Validation by gel electrophoresis

Use a Novex 6% polyacrylamide TBE pre-cast gel and 1X Novex TBE Running Buffer. For more information to do this step, consult the manufacturer's recommendations.

- Prepare an undigested DNA gel control by combining 2 µL of the Enrichment Control DNA stock solution and 2 µL of nuclease-free water.
- Add 1 µL of Novex Hi-Density TBE Sample Buffer (5X) to each 4-µL ECD sample.
- Load 5 µL of each sample on the gel. In one or more adjacent lanes, load 200 ng of a 50-bp DNA ladder.
- Run the gel at 210 V for approximately 15 minutes.
- Stain the gel in 3X GelRed Nucleic Acid Stain for 10 minutes, and visualize bands under UV radiation.

See Figure 6 for sample gel results.

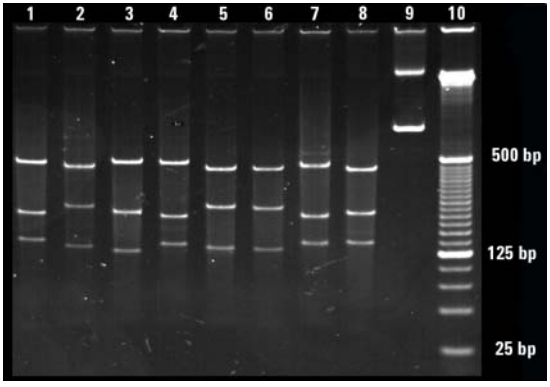


Figure 6 Validation of restriction digestion by gel electrophoresis. Lanes 1–8: ECD digestion reactions A–H, Lane 9: Undigested Enrichment Control DNA, Lane 10: 25-bp DNA ladder.

Stopping Point If you do not continue to the next step, samples may be stored at –20°C for long term storage. There are no more long-term stopping points until after the PCR amplification step on page 64.

Step 2. Hybridize digested DNA to HaloPlex or ClearSeq probe for target enrichment and sample indexing

In this step, the collection of gDNA restriction fragments is hybridized to the HaloPlex or ClearSeq probe capture library. HaloPlex and ClearSeq probes are designed to hybridize selectively to fragments originating from target regions of the genome and to direct circularization of the targeted DNA fragments. During the hybridization process, Illumina sequencing motifs including index sequences are incorporated into the targeted fragments.

The duration of the hybridization reaction is determined by the probe density of your design. Refer to the Certificate of Analysis provided with Box 1 of your kit to determine the hybridization conditions appropriate for your design.

For sample indexing primer assignments, see the Reference chapter for nucleotide sequences of the 96 indexes used in the HaloPlex Target Enrichment System.

CAUTION

This guide includes information for kits containing two different sets of indexing primers. **Verify that you are referencing the information appropriate for your kit version before you proceed.**

Kits with indexing primers supplied in a blue plate include 8-bp indexes A01 through H12. See page 87 through page 88 for indexing primer A01–H12 plate map and nucleotide sequence information.

Kits with indexing primers supplied in a clear plate include 8-bp indexes 1 through 96. See page 91 through page 97 for indexing primer 1–96 plate map and nucleotide sequence information.

Protocol steps for indexing using primers provided in either configuration are identical.

Prepare the NGS Bravo

1. Gently wipe down the Bravo deck with a NucleoClean decontamination wipe.
2. Place a red insert on Bravo deck position 1.
3. Place a silver Nunc plate insert on Bravo deck position 9.

4 Turn on the chiller, set to 0°C, at position 9 of the Bravo deck. Be sure that the chiller reservoir contains at least 300 mL of 25% ethanol.

5 For all run sizes, place a 384-well adapter insert on Bravo deck position 4. Pre-set the temperature of Bravo deck position 4 to 4°C using the Inheco Multi TEC control touchscreen, as described in Setting the Temperature of Bravo Deck Heat Blocks.

For 12-column runs only, place a second 384-well adapter insert on Bravo deck position 6 and pre-set the temperature of Bravo deck position 6 to 4°C.

NOTE

To expedite thermal cycler warm-up for the hybridization reaction on page 49, you can enter and initiate the hybridization program on the thermal cycler now, and then pause the program until you are instructed to transfer the reaction plate. Be sure to pause the thermal cycler before the initiation of the 95°C incubation segment. Release the pause immediately after transferring the plate to the thermal cycler in step 11 on page 49. Be sure that the 96-well block is in the thermal cycler before initiating the program for warm-up.

Prepare the Master Mix Source Plate for Hybridization.pro

1 Prepare the appropriate amount of Hybridization Master Mix, according to the table below.

Mix well by gentle vortexing, then spin the tube briefly.

Table 15 Preparation of Hybridization Master Mix for Hybridization.pro

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
HaloPlex or ClearSeq Probe	20 µL	255 µL	425 µL	595 µL	765 µL	1105 µL	2210 µL
Hybridization Solution	50 µL	637.5 µL	1062.5 µL	1487.5 µL	1912.5 µL	2762.5 µL	5525 µL
Total Volume	**70 µL**	**892.5 µL**	**1487.5 µL**	**2082.5 µL**	**2677.5 µL**	**3867.5 µL**	**7735 µL**

2 In a Nunc DeepWell plate, prepare the Hybridization Master Mix source plate. Add the volumes indicated in Table 16 of the Hybridization Master Mix to all wells of the indicated column of the Nunc DeepWell plate.

Table 16 Preparation of the Master Mix Source Plate for Hybridization.pro

Master Mix Solution	Position on Source Plate	Volume of Master Mix added per Well of Nunc DeepWell Source Plate					
		1-Column Runs	2-Column Runs	3-Column Runs	4-Column Runs	6-Column Runs	12-Column Runs
Hybridization Master Mix	Column 1 (A1-H1)	102.8 µL	177.2 µL	251.6 µL	325.9 µL	474.7 µL	958.1 µL

Prepare the Hybridization Reaction Plate with Indexing Primers

CAUTION

This guide includes information for kits containing two different sets of indexing primers. **Verify that you are referencing the information appropriate for your kit version before you proceed.**

Reference information for indexing primers supplied in a blue plate begins on page 84.

Reference information for indexing primers supplied in a clear plate begins on page 89.

Protocol steps for indexing using primers provided in either configuration are identical.

1 In a half-skirted 96-well Eppendorf twin.tec plate, aliquot 10 µL of the appropriate HaloPlex Indexing Primer to each intended sample indexing well position. Keep the plate on ice.

Be sure to add only one specific Indexing Primer to each well, using different indexes for each sample to be multiplexed. Record the identity of the index assigned to each well for later sequence analysis.

2 If the run includes an ECD control sample that was analyzed as described on page 39, add 32 µL of nuclease-free water to well A1 of the hybridization reaction plate. (Well A1 should also contain 10 µL of indexing primer from step 1 above.) The 32 µL of water added-back here compensates for the combined volume removed from the eight ECD digest wells during validation.

Load the Agilent NGS Bravo and Run the Hybridization.pro VWorks Protocol

1 On the VWorks HaloPlex form, under **Step**, select **02 Hybridization.pro.**

2 Select the number of columns of samples to be processed. Runs must include 1, 2, 3, 4, 6, or 12 columns.

3 Click **Update layout and information.**

4 Load the Bravo deck according to Table 17.

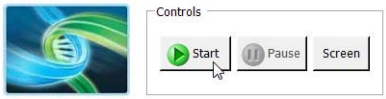
Table 17 Initial Bravo deck configuration for Hybridization.pro

Location	Content
1	HaloPlex Indexing Primer source plate (half-skirted 96-well Eppendorf twin.tec plate) seated on red insert
2	New tip box
3	—(empty)—
4	Digested DNA in 384-well plate, seated on 384-well insert
5	Empty full-skirted 96-well Eppendorf twin.tec plate
6	For 12-column runs only: Digested DNA in 384-well plate (digest plate 2), seated on 384-well insert For 1- to 6-column runs: empty
7	—(empty)—
8	Empty tip box
9	Hybridization Master Mix source plate (Nunc DeepWell plate) seated on silver insert

5 Verify that the NGS Bravo has been set up as displayed in the **Bravo Deck Setup** and **Information** regions of the form.

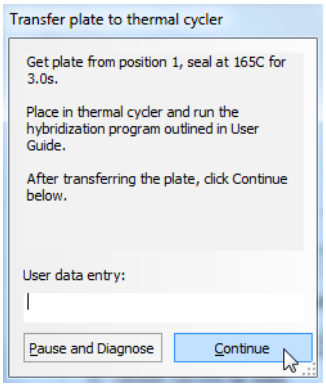
6 Verify that the **Current Tip State** indicator on the form matches the configuration of unused and used tips in the tip boxes at Bravo Deck positions 2 and 8, respectively. See page 21 for more information on using this segment of the form during the run.

7 When verification is complete, click **Start** to start the run.



The NGS Bravo combines all eight digestion reactions for each gDNA sample with Hybridization Master Mix and the appropriate Indexing Primer in wells of a 96-well plate.

8 When the NGS Bravo has finished preparing the hybridization plate for the run, you will be prompted by VWorks as shown below.


Transfer plate to thermal cycler
Get plate from position 1, seal at 165C for 3.0s.
Place in thermal cycler and run the hybridization program outlined in User Guide.
After transferring the plate, click Continue below.
User data entry:
Pause and Diagnose
Continue

9 Seal the sample plate using the PlateLoc Thermal Microplate Sealer, with sealing settings of 165°C and 3.0 sec.

10 Spin the plate briefly.

11 Transfer the sealed plate to a thermal cycler and run the appropriate program in Table 18, using the hybridization duration listed on the Certificate of Analysis. After transferring the plate, click **Continue** on the VWorks prompt to finish the protocol.

Use a heated lid. Do **not** include a low-temperature hold step in the thermal cycler program. Incubation at 54°C for more than the indicated time is not recommended.

Table 18 Thermal cycler program* for probe hybridization

Step	Temperature	Time (Duration of Step)	
		Designs with <20,000 probes (see Certificate of Analysis)	Designs with >20,000 probes (see Certificate of Analysis)
Step 1	95°C	10 minutes	10 minutes
Step 2	54°C	3 hours	16 hours

* Thermal cyclers that use calculated temperature methods cannot be set to 160 µL reaction volumes. In that case, enter the maximum possible volume.

CAUTION

Make sure that the thermal cycler has a maximum reaction volume specification of at least 100 µL.

The 160-µL hybridization reaction conditions have been optimized with the SureCycler thermal cycler (with volume specification of 10-100 µL for PCR reactions). The performance of other thermal cyclers for this application should be verified before use.

Step 3. Capture and amplify the target DNA

In this step, the circularized target DNA-probe hybrids, containing biotin, are captured on streptavidin beads. After capture, DNA ligase is added to seal nicks, then target DNA is eluted and PCR-amplified.

Assemble reagents for the run

1 Remove reagents to be used in upcoming protocol steps from cold storage and allow the solutions to reach room temperature:

- From –20°C storage, remove the Capture Solution, Wash Solution, Ligation Solution and SSC Buffer.
- From +4°C storage, remove the HaloPlex Magnetic Beads.

2 Prepare 30 µL per sample, plus excess, of fresh 50 mM NaOH for use in the DNA elution step on page 52.

Prepare the 50 mM NaOH solution from a 10M NaOH stock solution.

CAUTION

Using high-quality NaOH is critical for optimal DNA elution and recovery.

- Do not use stock NaOH solutions that were stored at concentrations below 10 M to prepare the 50 mM NaOH solution.
- Keep the 50 mM NaOH solution container sealed when not in use, especially when processing large numbers of samples per run.

Table 19 Amount of 50mM NaOH required per run size

Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
30 µL	270 µL	510 µL	750 µL	990 µL	1470 µL	2940 µL

3 Obtain or prepare 0.5 µL per sample, plus excess, of 2 M acetic acid, for use in the PCR master mix on page 53.

CAUTION

It is critical to use high-quality acetic acid at 2 M concentration in this step to ensure neutralization of the NaOH used for elution.

See Table 1 on page 9 for 2 M acetic acid supplier information, or prepare 2 M acetic acid from high-quality glacial acetic acid.

Prepare the NGS Bravo

1 Gently wipe down the Bravo deck with a NucleoClean decontamination wipe.

2 Place a red insert on Bravo deck position 4.

3 Pre-set the temperature of Bravo deck position 4 to 54°C using the Inheco Multi TEC control touchscreen, as described in Setting the Temperature of Bravo Deck Heat Blocks.

4 Place a second red insert on Bravo deck position 6.

5 Pre-set the temperature of Bravo deck position 6 to 4°C using the Inheco Multi TEC control touchscreen, as described in Setting the Temperature of Bravo Deck Heat Blocks.

6 Place the silver Nunc plate insert on Bravo deck position 9.

7 Turn on the chiller, set to 0°C, at position 9 of the Bravo deck. Be sure that the chiller reservoir contains at least 300 mL of 25% ethanol.

Prepare the HaloPlex Magnetic Beads Source Plate

1 Vigorously resuspend the HaloPlex Magnetic Beads on a vortex mixer. The beads settle during storage.

2 Wash the magnetic beads.

a Transfer 40 µL per sample of the HaloPlex Magnetic Beads suspension to a 1.5-mL tube or conical vial, using volumes provided in Table 20.

Table 20 Volume of HaloPlex Magnetic Bead suspension for capture

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
HaloPlex Magnetic Beads	0.04 mL	0.36 mL	0.68 mL	1.0 mL	1.32 mL	1.96 mL	3.92 mL

b Put the vial into a compatible magnetic device for 5 minutes.

c After verifying that the solution has cleared, carefully remove and discard the supernatant using a pipette.

d Add an equivalent volume of Capture Solution (see Table 21) to the beads and resuspend by pipetting up and down.

Table 21 Volume of Capture Solution used for bead resuspension

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
Capture Solution	0.04 mL	0.36 mL	0.68 mL	1.0 mL	1.32 mL	1.96 mL	3.92 mL

3 Prepare a Nunc DeepWell source plate for the washed HaloPlex streptavidin bead suspension. Add 40 µL of the homogeneous bead suspension to all wells of the Nunc DeepWell plate that correspond to sample-containing wells on the hybridization plate.

4 Place the streptavidin bead source plate at position 5 of the Bravo deck.

Prepare wash and elution solution source plates

Prepare a separate source plate for each of the solutions listed in Table 22. Use full-skirted 96-well Eppendorf twin.tec plates to prepare all three source plates. For all sample-containing wells of the hybridization plate, add the specified volume of solution to all corresponding wells of the solution source plate.

Table 22 Preparation of solution source plates for Capture_v1.1.pro protocol

Solution	Volume to dispense per well of source plate
Wash Solution	110 µL
SSC Buffer	110 µL
50 mM NaOH	30 µL

Seal the 50 mM NaOH source plate using the PlateLoc Thermal Microplate Sealer, with sealing settings of 165°C and 1.0 sec. Leave sealed until you are prompted to add the plate to the Bravo deck in step 15 on page 62.

Prepare the Master Mixes for Capture_v1.1.pro protocol

1 Prepare the appropriate amount of PCR Master Mix, according to the table below.

Mix well by gentle vortexing, then spin the tube briefly.

Table 23 Preparation of PCR Master Mix for Capture_v1.1.pro

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
Nuclease-free water	16.1 µL	205.3 µL	342.1 µL	479 µL	615.8 µL	889.5 µL	1779 µL
5X Herculase II Reaction Buffer	10 µL	127.5 µL	212.5 µL	297.5 µL	382.5 µL	552.5 µL	1105 µL
dNTPs (100 mM)*	0.4 µL	5.1 µL	8.5 µL	11.9 µL	15.3 µL	22.1 µL	44.2 µL
Primer 1	1 µL	12.75 µL	21.3 µL	29.8 µL	38.3 µL	55.3 µL	110.5 µL
Primer 2	1 µL	12.75 µL	21.3 µL	29.8 µL	38.3 µL	55.3 µL	110.5 µL
2 M Acetic acid	0.5 µL	6.4 µL	10.6 µL	14.9 µL	19.1 µL	27.6 µL	55.3 µL
Herculase II Fusion DNA Polymerase	1 µL	12.75 µL	21.3 µL	29.8 µL	38.3 µL	55.3 µL	110.5 µL
Total Volume	**30 µL**	**382.5 µL**	**637.6 µL**	**892.7 µL**	**1147.6 µL**	**1657.6 µL**	**3315 µL**

* Be sure to use dNTPs at 100 mM concentration (25 mM for each nucleotide), like those provided with the Herculase II Fusion Enzyme with dNTPs (Agilent p/n 600677 or 600679).

2 Prepare the appropriate amount of Ligation Master Mix, according to the table below.

Mix well by gentle vortexing, then spin the tube briefly. Store the master mix on ice until it is used on page 58. The Ligation Master Mix is added to the Master Mix Source Plate just before it is used in the Capture_v1.1.pro protocol. Do not add this master mix to the source plate before starting the run.

Table 24 Preparation of Ligation Master Mix for Capture_v1.1.pro

Reagent	Volume for 1 Library	Volume for 1 Column	Volume for 2 Columns	Volume for 3 Columns	Volume for 4 Columns	Volume for 6 Columns	Volume for 12 Columns
Ligation Solution	47.5 µL	605.6 µL	1009 µL	1413 µL	1817 µL	2624 µL	5249 µL
DNA Ligase	2.5 µL	31.9 µL	53.1 µL	74.4 µL	95.6 µL	138.1 µL	276.3 µL
Total Volume	**50 µL**	**637.5 µL**	**1062.1 µL**	**1487.4 µL**	**1912.6 µL**	**2762.1 µL**	**5525.3 µL**

Prepare the Master Mix Source Plate for Capture_v1.1.pro

Using the same Nunc DeepWell plate that was used for the Hybridization.pro run, prepare the Master Mix source plate for Capture_v1.1.pro. Add the volume indicated in Table 25 of PCR Master Mix to all wells of column 3 of the Nunc DeepWell plate.

Table 25 Preparation of the Master Mix Source Plate for Capture_v1.1.pro

Master Mix Solution	Position on Source Plate	Volume of Master Mix added per Well of Nunc Deep Well Source Plate					
		1-Column Runs	2-Column Runs	3-Column Runs	4-Column Runs	6-Column Runs	12-Column Runs
PCR Master Mix	Column 3 (A3-H3)	44.1 µL	75.9 µL	107.8 µL	139.7 µL	203.4 µL	410.6 µL

NOTE

Column 2 of the Master Mix source plate must remain empty at this step. You will be prompted to add Ligation Master Mix to Column 2 at the appropriate time during the Capture_v1.1.pro protocol. Column 1 was used during the Hybridization.pro protocol.

If you are using a new DeepWell plate for the Capture_v1.1.pro Master Mix source plate, be sure to leave columns 1 and 2 empty at this time, adding the PCR Master Mix to column 3 of the new plate.

Load the Agilent NGS Bravo and Run the Capture_v1.1.pro VWorks Protocol

1 On the VWorks HaloPlex form, under **Step**, select **03 Capture_v1.1.pro.**

2 Select the number of columns of samples to be processed. Runs must include 1, 2, 3, 4, 6, or 12 columns.

3 Click **Update layout and information.**

4 Load the Bravo deck according to Table 26.

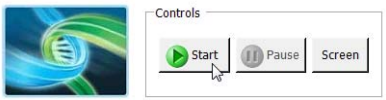
Table 26 Initial Bravo deck configuration for Capture_v1.1.pro

Location	Content
1	Empty Axygen 96 Deep Well Plate (square wells) for waste
2	New tip box
3	Wash Solution source plate (full-skirted 96-well Eppendorf twin.tec plate)
4	Hybridized sample plate seated on red insert
5	HaloPlex magnetic streptavidin bead source plate (Nunc DeepWell plate)
6	Empty half-skirted 96-well Eppendorf twin.tec plate seated on red insert
7	—(empty)—
8	Empty tip box
9	Master Mix source plate (Nunc DeepWell plate) seated on silver insert

5 Verify that the NGS Bravo has been set up as displayed in the **Bravo Deck Setup** and **Information** regions of the form.

6 Verify that the **Current Tip State** indicator on the form matches the configuration of unused and used tips in the tip boxes at Bravo Deck positions 2 and 8, respectively. See page 21 for more information on using this segment of the form during the run.

7 When verification is complete, click **Start** to start the run.

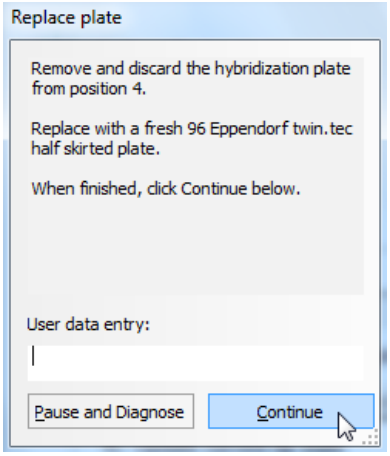


The NGS Bravo completes the liquid-handling steps for capture of the target DNA-HaloPlex probe hybrids on the streptavidin beads.

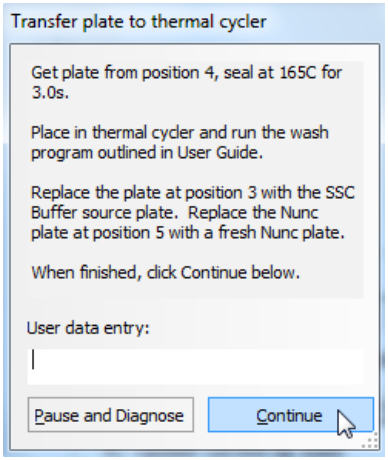
NOTE

To expedite thermal cycler warm-up for the subsequent wash program on page 57, you can enter and initiate the wash program on the thermal cycler now, and then pause the program until you are instructed to transfer the reaction plate. Be sure to pause the thermal cycler before the initiation of the 46°C incubation segment. Release the pause immediately after transferring the plate to the thermal cycler in step 9.

8 When prompted by VWorks as shown below, remove and discard the hybridization plate from position 4 of the Bravo deck. Place a fresh half-skirted 96-well Eppendorf twin.tec plate at position 4 for use in the wash segment of the protocol.


Replace plate
Remove and discard the hybridization plate from position 4.
Replace with a fresh 96 Eppendorf twin.tec half skirted plate.
When finished, click Continue below.
User data entry:
Pause and Diagnose
Continue

9 When the NGS Bravo has finished preparing the capture wash plate, you will be prompted by VWorks as shown below.


Transfer plate to thermal cycler
Get plate from position 4, seal at 165C for 3.0s.
Place in thermal cycler and run the wash program outlined in User Guide.
Replace the plate at position 3 with the SSC Buffer source plate. Replace the Nunc plate at position 5 with a fresh Nunc plate.
When finished, click Continue below.
User data entry:
Pause and Diagnose
Continue

a Get the sample plate from position 4 and seal the plate using the PlateLoc Thermal Microplate Sealer, with sealing settings of 165°C and 3.0 sec.

Transfer the sealed plate to a thermal cycler and run the wash program shown in Table 27, using a heated lid.

Do **not** include a low-temperature hold step in the thermal cycler program following the 10-minute incubation.

Table 27 Thermal cycler program for Capture_v1.1.pro wash step

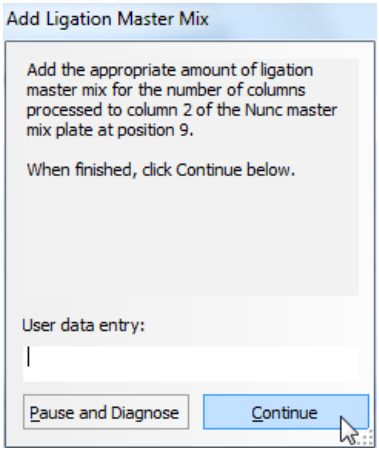
Step	Temperature	Time
Step 1	46°C	10 minutes

b Remove and discard the Wash Solution plate from position 3 of the Bravo deck. Place the SSC Buffer source plate at position 3 for use in the ligation segment of the protocol.

c Remove and discard the bead source plate from position 5 of the Bravo deck. Place a fresh Nunc DeepWell plate at position 5.

d After completing all steps, click **Continue** on the VWorks prompt to continue the automation protocol. Do not wait for the conclusion of the thermal cycler wash program to continue the protocol.

10 During the 10-minute incubation of the sample plate on the thermal cycler, you will be prompted to add the Ligation Master Mix to the Master Mix source plate as shown below.


Add Ligation Master Mix
Add the appropriate amount of ligation master mix for the number of columns processed to column 2 of the Nunc master mix plate at position 9.
When finished, click Continue below.
User data entry:
Pause and Diagnose
Continue

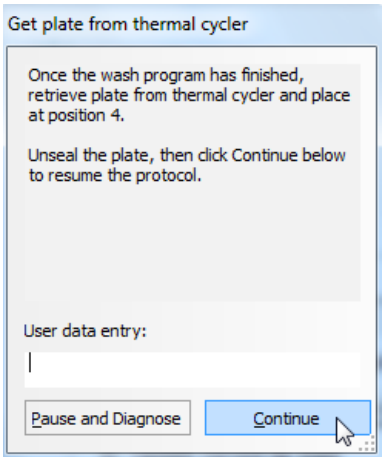
Add the volume of Ligation Master Mix indicated in Table 28 to all wells of column 2 of the Nunc DeepWell Master Mix source plate on Bravo deck position 9.

Table 28 Addition of Ligation Master Mix to the Master Mix Source Plate for Capture_v1.1.pro

Master Mix Solution	Position on Source Plate	Volume of Master Mix added per Well of Nunc Deep Well Source Plate					
		1-Column Runs	2-Column Runs	3-Column Runs	4-Column Runs	6-Column Runs	12-Column Runs
Ligation Master Mix	Column 2 (A2-H2)	73.4 µL	126.6 µL	179.7 µL	232.8 µL	339.1 µL	684.4 µL

NOTE The Master Mix source plate at postition 9 should already contain the PCR Master Mix in Column 3 and the depleted Hybridization Master Mix from the Hybridization.pro protocol in Column 1. Be sure to add the Ligation Master Mix to Column 2 of the source plate at this step.

11 Once the wash program in Table 27 has finished and you are prompted by VWorks, transfer the plate from the thermal cycler to Bravo deck position 4. Carefully unseal the plate, then click Continue on the VWorks prompt to resume the Capture_v1.1.pro protocol.

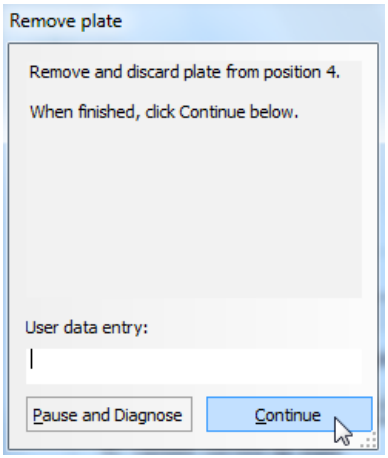

Get plate from thermal cycler
Once the wash program has finished, retrieve plate from thermal cycler and place at position 4.
Unseal the plate, then click Continue below to resume the protocol.
User data entry:
Pause and Diagnose
Continue

The NGS Bravo completes the liquid-handling steps for ligation of the captured target DNA.

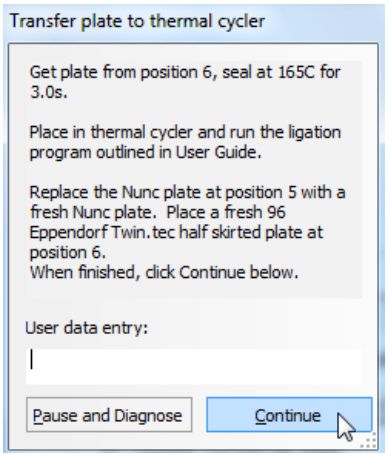
NOTE

To expedite thermal cycler warm-up for the subsequent ligation program on page 61, you can enter and initiate the wash program on the thermal cycler now, and then pause the program until you are instructed to transfer the reaction plate. Be sure to pause the thermal cycler before the initiation of the 55°C incubation segment. Release the pause immediately after transferring the plate to the thermal cycler in step 13.

12 When prompted by VWorks as shown below, remove and discard the plate from position 4. Click Continue on the prompt to resume the protocol.



13 When the NGS Bravo has finished preparing the ligation plate, you will be prompted by VWorks as shown below.


Transfer plate to thermal cycler
Get plate from position 6, seal at 165C for 3.0s.
Place in thermal cycler and run the ligation program outlined in User Guide.
Replace the Nunc plate at position 5 with a fresh Nunc plate. Place a fresh 96 Eppendorf Twin.tec half skirted plate at position 6.
When finished, click Continue below.
User data entry:
Pause and Diagnose
Continue

a Get the sample plate from position 6 and seal the plate using the PlateLoc Thermal Microplate Sealer, with sealing settings of 165°C and 3.0 sec.

b Transfer the sealed plate to a thermal cycler and run the ligation program shown in Table 29, using a heated lid.

Table 29 Thermal cycler program for Capture_v1.1.pro ligation step

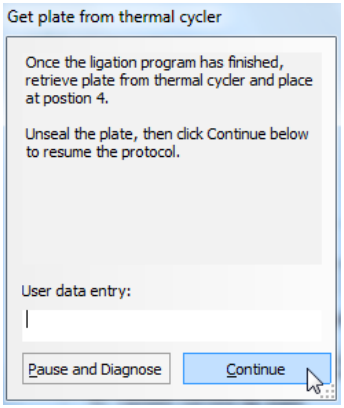
Step	Temperature	Time
Step 1	55°C	10 minutes
Step 2	4°C	Hold

c Remove and discard the Nunc DeepWell plate from position 5 of the Bravo deck. Place a fresh Nunc DeepWell plate at position 5 for use in the ligation purification steps of the protocol.

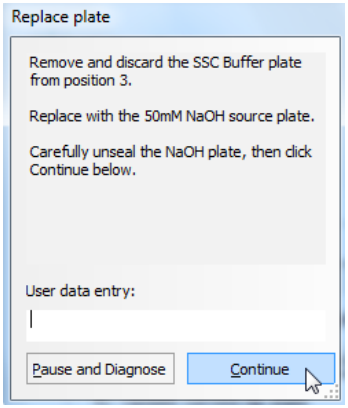
d Place a fresh half-skirted 96-well Eppendorf twin.tec plate at position 6 for use in the following PCR protocol steps.

e After completing all steps, click **Continue** on the VWorks prompt to continue the automation protocol. Do not wait for the conclusion of the thermal cycler ligation program to continue the protocol.

14 Once the ligation program in Table 29 is finished and you are prompted by VWorks, transfer the plate from the thermal cycler to Bravo deck position 4. Carefully unseal the plate, then click **Continue** on the VWorks prompt to resume the Capture_v1.1.pro protocol.


Get plate from thermal cycler
Once the ligation program has finished, retrieve plate from thermal cycler and place at postion 4.
Unseal the plate, then click Continue below to resume the protocol.
User data entry:
Pause and Diagnose
Continue

15 When prompted by VWorks as shown below, remove and discard the SSC Buffer plate from position 3 of the Bravo deck. Place the 50 mM NaOH source plate at position 3 for use in the elution steps of the protocol. After carefully unsealing the source plate, click **Continue** on the prompt to resume the protocol.

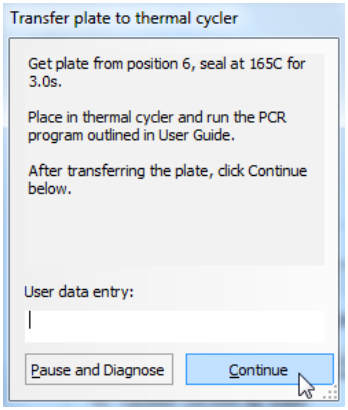

Replace plate
Remove and discard the SSC Buffer plate from position 3.
Replace with the 50mM NaOH source plate.
Carefully unseal the NaOH plate, then click Continue below.
User data entry:
Pause and Diagnose
Continue

The NGS Bravo completes the liquid-handling steps for elution of the captured target DNA followed by preparation of PCR reactions for amplification.

NOTE

To expedite thermal cycler warm-up for the subsequent PCR program on page 64, you can enter and initiate the PCR program on the thermal cycler now, and then pause the program until you are instructed to transfer the reaction plate. Be sure to pause the thermal cycler before the initiation of the 98°C denaturation segment. Release the pause immediately after transferring the plate to the thermal cycler in step 16.

16 When the NGS Bravo has finished preparing the PCR amplification reactions, you will be prompted by VWorks as shown below.


Transfer plate to thermal cycler
Get plate from position 6, seal at 165C for 3.0s.
Place in thermal cycler and run the PCR program outlined in User Guide.
After transferring the plate, click Continue below.
User data entry:
Pause and Diagnose
Continue

a Get the sample plate from position 6 and seal the plate using the PlateLoc Thermal Microplate Sealer, with sealing settings of 165°C and 3.0 sec.

b Transfer the sealed plate to a thermal cycler and run the PCR program in Table 30, using a heated lid.

The optimal amplification cycle number varies for each HaloPlex or ClearSeq Probe design. Consult the Certificate of Analysis (provided with HaloPlex Target Enrichment System Box 1) for the PCR cycling recommendation for your probe.

Table 30 HaloPlex post-capture DNA amplification PCR program

Segment	Number of Cycles	Temperature	Time
1	1	98°C	2 minutes
2	Obtain cycle number from Certificate of Analysis	98°C	30 seconds
		60°C	30 seconds
		72°C	1 minute
3	1	72°C	10 minutes
4	1	8°C	Hold

c After initiating the PCR program in the thermal cycler, click **Continue** on the VWorks prompt to finish the automation protocol.

d If you are continuing to the next step of PCR product purification, remove the Agencourt AMPure XP Beads from +4°C storage for use on page 65. Let the beads come to room temperature for the remainder of the amplification program.

Stopping Point If you do not continue to the next step, PCR products may be stored at –20°C for up to 72 hours or at 8°C overnight. For best results, however, purify PCR products as soon as possible.

Step 4. Purify the amplified target DNA

In this step, the NGS Bravo does the liquid handling steps to purify the amplified target DNA sample using AMPure XP beads.

Prepare the NGS Bravo and reagents

1. Gently wipe down the Bravo deck with a NucleoClean decontamination wipe.
2. Let the AMPure XP beads come to room temperature for at least 30 minutes.

   Do not freeze the AMPure XP beads at any time.
3. Mix the room-temperature bead suspension well so that the reagent appears homogeneous and consistent in color.
4. Prepare a Nunc DeepWell source plate containing AMPure XP beads. For each well to be processed, add 100 µL of homogeneous AMPure XP beads per well to the Nunc DeepWell plate.
5. Place a red insert on Bravo deck position 6.
6. Pre-set the temperature of Bravo deck position 6 to 4°C using the Inheco Multi TEC control touchscreen, as described in Setting the Temperature of Bravo Deck Heat Blocks.
7. Prepare a Thermo Scientific reservoir containing 15 mL of nuclease-free water.
8. Prepare a Thermo Scientific reservoir containing 15 mL of the final sample elution buffer [nuclease-free 10 mM Tris-acetate or Tris-HCl buffer (pH 8.0)].
9. Prepare a separate Thermo Scientific reservoir containing 45 mL of freshly-prepared 70% ethanol.

Load the Agilent NGS Bravo and Run the Purification_v1.1.pro VWorks Protocol

1. On the VWorks HaloPlex form, under **Step**, select **04 Purification_v1.1.pro.**
2. Select the number of columns of samples to be processed. Runs must include 1, 2, 3, 4, 6, or 12 columns.
3. Click **Update layout and information.**

4 Load the Bravo deck according to Table 31.

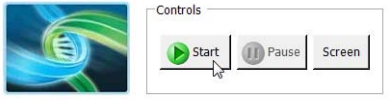
Table 31 Initial Bravo deck configuration for Purification_v1.1.pro

Location	Content
1	Empty Axygen 96 Deep Well Plate (square wells) for waste
2	New tip box
3	Empty full-skirted 96-well Eppendorf twin.tec plate
4	—(empty)—
5	AMPure XP beads in Nunc DeepWell source plate
6	Amplified DNA samples in half-skirted 96-well Eppendorf twin.tec plate seated on red insert
7	—(empty)—
8	Empty tip box
9	Nuclease-free water in Thermo Scientific reservoir

5 Verify that the NGS Bravo has been set up as displayed in the **Bravo Deck Setup** region of the form.

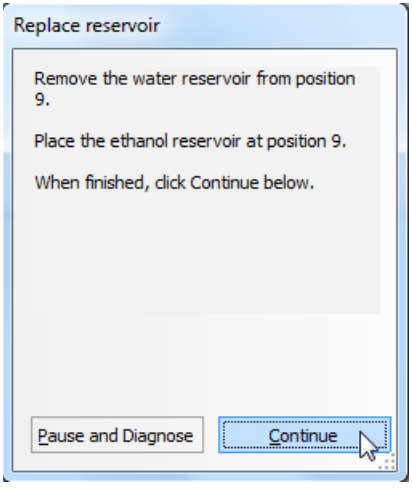
6 Verify that the **Current Tip State** indicator on the form matches the configuration of unused and used tips in the tip boxes at Bravo Deck positions 2 and 8, respectively. See page 21 for more information on using this segment of the form during the run.

7 When verification is complete, click **Start** to start the run.



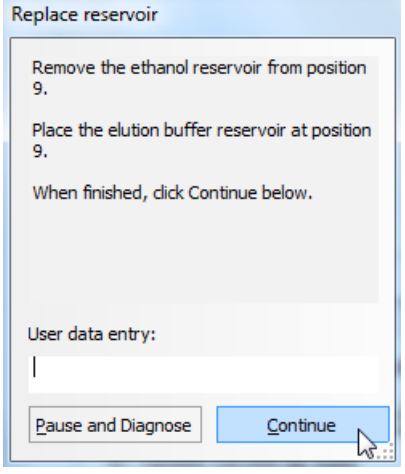
8 When prompted by VWorks as shown below, remove the water reservoir from position 9 of the Bravo deck and replace it with the 70% ethanol reservoir.

When finished, click **Continue** on the VWorks prompt.


Replace reservoir
Remove the water reservoir from position 9.
Place the ethanol reservoir at position 9.
When finished, click Continue below.
Pause and Diagnose
Continue

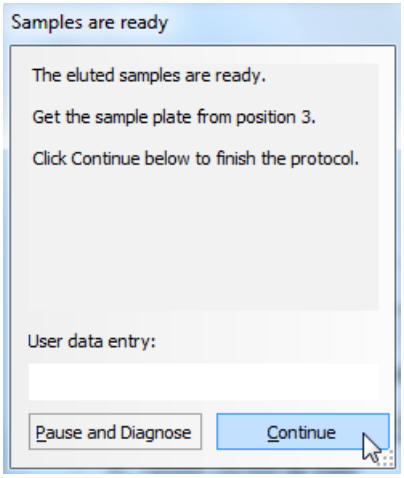
9 When prompted by VWorks as shown below, remove the 70% ethanol reservoir from position 9 of the Bravo deck and replace it with the final sample elution buffer reservoir.

When finished, click **Continue** on the VWorks prompt.


Replace reservoir
Remove the ethanol reservoir from position 9.
Place the elution buffer reservoir at position 9.
When finished, click Continue below.
User data entry:
Pause and Diagnose
Continue

The NGS Bravo completes the liquid-handling steps for elution of the captured target DNA.

10 When the NGS Bravo has finished preparing the final eluted sample plate, you will be prompted by VWorks as shown below. Click **Continue** on the VWorks prompt to finish the protocol.


Samples are ready
The eluted samples are ready.
Get the sample plate from position 3.
Click Continue below to finish the protocol.
User data entry:
Pause and Diagnose
Continue

Stopping Point If you do not continue to the next step, samples may be stored at –20°C for long-term storage (up to one year). Avoid subjecting the stored DNA samples to multiple freeze-thaw cycles.

Step 5. Validate enrichment and quantify enriched target DNA

Prior to sample pooling and sequencing sample preparation, validate enrichment and quantify the enriched target DNA in each library sample by microfluidic analysis using the 2100 Bioanalyzer (see page 70) or the 2200 TapeStation (see page 71).

Enriched library samples may also be qualitatively analyzed using gel electrophoresis. Sample gel electrophoresis results are provided in the Reference section on page 98.

Expected Results

Each amplicon in the prepared library contains one target insert surrounded by sequence motifs required for multiplexed sequencing using the Illumina platform. Amplicons include 50 to 500 bp of target DNA insert and 125 bp of sequencing motifs, as shown in Figure 7.

Figure 7 Content of target-enriched amplicons. Each amplicon contains one target insert (blue) surrounded by the Illumina paired-end sequencing elements (black), the sample index (red) and the library bridge PCR primers (yellow).

The amplicons should range from 175 to 625 bp in length, with the majority of products sized 225 to 525 bp. Amplicons in the 175 to 625 bp size range should be included for quantitation of the enriched target DNA in each sample. Any spurious DNA products outside of this size range in any sample should be excluded from the target DNA quantitation results.

Option 1: Analysis using the 2100 Bioanalyzer System

Use a Bioanalyzer High Sensitivity DNA Assay kit and the 2100 Bioanalyzer instrument with 2100 Expert Software (version B.02.07 or higher required to run the High Sensitivity Kit). See the reagent kit guide for general Bioanalyzer instrument and assay setup instructions.

1 Prepare the chip, samples and ladder as instructed in the reagent kit guide, using 1 µL of enriched library sample for the analysis.

2 Load the prepared chip into the 2100 Bioanalyzer instrument and start the run within five minutes after preparation.

3 Analyze the electropherogram for each sample according to the analysis guidelines on page 72.

See Figure 8 for a sample Bioanalyzer electropherogram.

NOTE

If the concentration determined by Bioanalyzer analysis is > 10 ng/µL, repeat the analysis using a 1:10 dilution of the sample. Dilute 1 µL of the sample in 9 µL of 10 mM Tris, 1 mM EDTA and then mix well by vortexing at 2000 rpm on the IKA vortex supplied with the Bioanalyzer before analyzing the diluted sample.

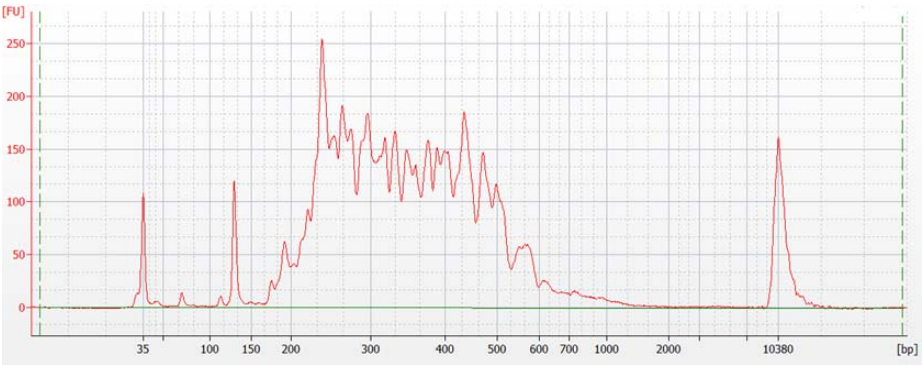


Figure 8 Validation of HaloPlex enrichment by 2100 Bioanalyzer system analysis.

Option 2: Analysis using the 2200 TapeStation

Use a High Sensitivity D1000 ScreenTape and reagent kit. For more information to do this step, see the *Agilent 2200 TapeStation User Manual* at www.genomics.agilent.com.

1 Prepare the TapeStation samples as instructed in the *2200 TapeStation User Manual*. Use 2 µL of each enriched library sample diluted with 2 µL of High Sensitivity D1000 sample buffer in separate wells of a tube strip for the analysis.

2 Load the sample tube strip, the High Sensitivity D1000 ScreenTape, and loading tips into the 2200 TapeStation as instructed in the *2200 TapeStation User Manual*. Start the run.

3 Analyze the electropherogram for each sample according to the analysis guidelines on page 72.

See Figure 9 for a sample TapeStation electropherogram.

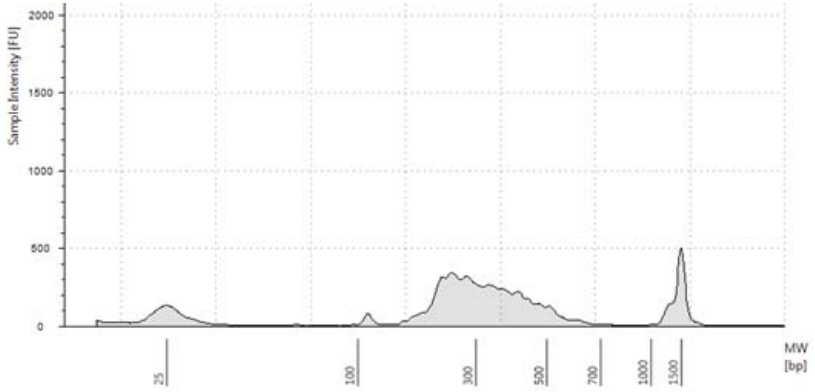


Figure 9 Validation of HaloPlex enrichment by 2200 TapeStation analysis.

Analysis of Electropherogram Results

- Check that the electropherogram shows a peak fragment size between approximately 225 to 525 bp.
- Determine the concentration of enriched target DNA in the sample by integration under the peak between 175 and 625 bp. Peaks at <150 bp may be observed, but should be excluded from quantitation.
- Some designs may generate a peak at about 125 bp. This peak is associated with an adaptor-dimer product which will cluster and generate sequence that does not map to the genome. If the molar fraction of the 125 bp peak is greater than 10%, do another round of AMPure purification after pooling samples. First, pool equimolar amounts of libraries to be multiplexed, using concentrations determined for the 175–625 peak of each sample. Using 40 µL of the pooled libraries, purify the DNA using AMPure XP beads according to the protocol starting on page 79.

Step 6. Pool samples with different indexes for multiplexed sequencing

Use the following guidelines to design your sample pooling strategy:

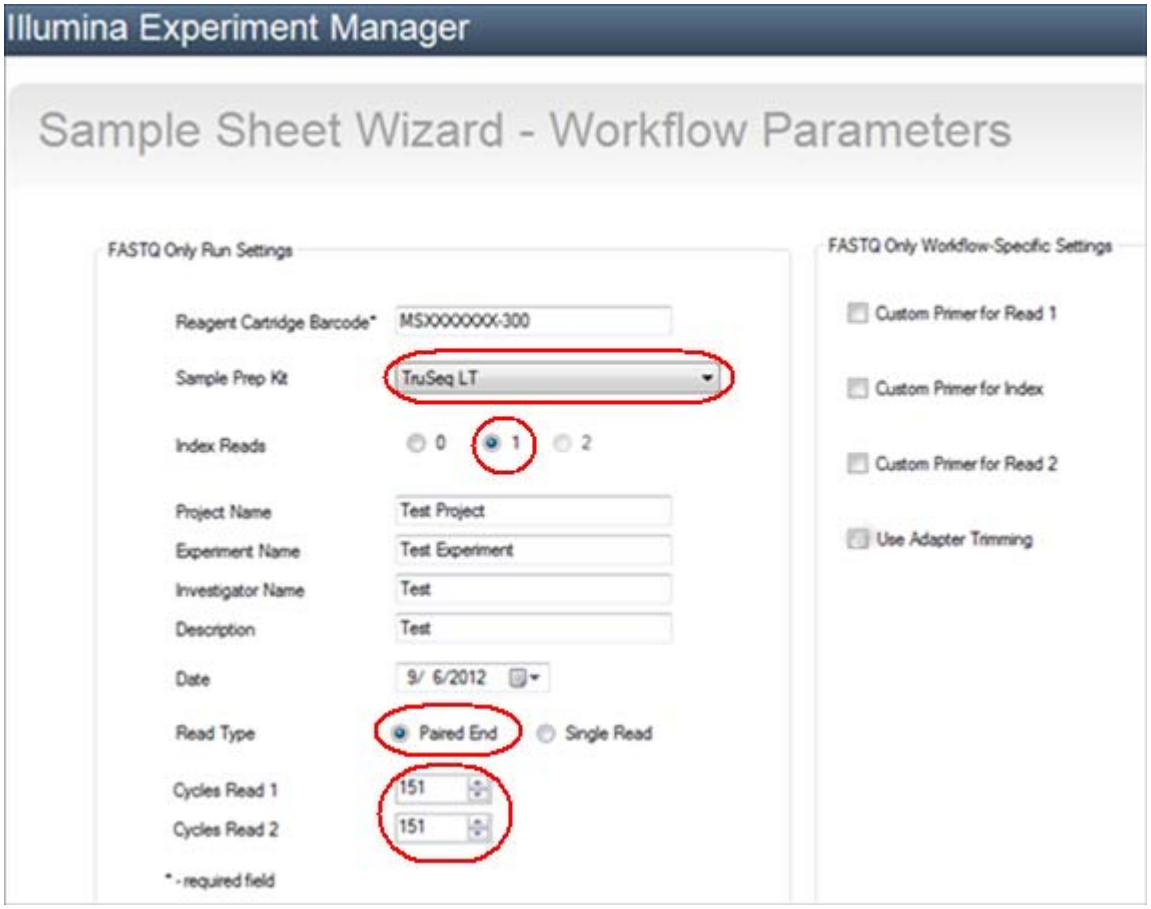
- Use the Bioanalyzer- or TapeStation-measured concentration of 175-625 bp products in each sample to pool equimolar amounts of differentially indexed samples in order to optimize the use of sequencing capacity.
- The final HaloPlex enrichment pool is ready for direct sequencing using standard Illumina paired-end primers and chemistry on the Illumina HiSeq, MiSeq, or GAIIx platform. See additional guidelines for the MiSeq platform (below) and HiSeq platform (page 77).
- Use 100 + 100 bp or 150 + 150 bp paired-end sequencing, depending on the selection made during probe design. Since the read length affects maximum achievable coverage, check the design report to verify read length selected in probe design.
- Sequencing runs must be set up to perform an 8-nt index read. For complete index sequence information, see the Reference chapter starting on page 83.
- Before aligning reads to the reference genome, trim the reads from Illumina adaptor sequences.

MiSeq platform sequencing run setup guidelines

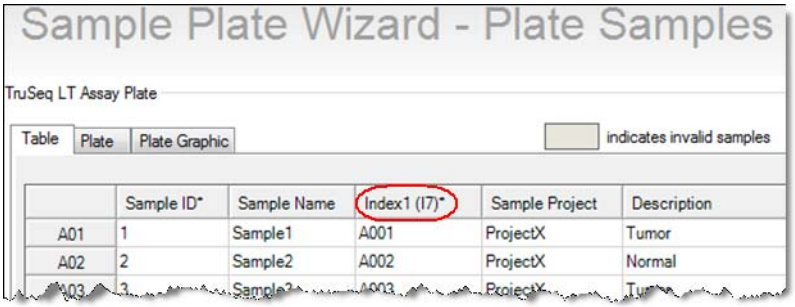
Setting up a custom Sample Sheet:

1 In the IEM software, create a Sample Sheet for the MiSeq platform using the following Workflow selections.
   - Under **Category**, select *Other*.
   - Under **Application**, select *FASTQ Only*.

2 On the **Workflow Parameters** screen, enter the run information, making sure to specify the key parameters highlighted below:



3 Using the **Sample Plate Wizard**, set up a New Plate, entering the required information for each sample to be sequenced. In the **Index 1(17)** column of the **TrueSeq LT Assay Plate** table, assign each sample to any of the Illumina 17 indexes. The index will be corrected to a HaloPlex index at a later stage.



4 Finish the sample plate setup tasks and save the sample plate file.

5 Using the **Sample Sheet Wizard**, select the samples to include in the run and save the Sample Sheet file.

Editing the Sample Sheet to Include HaloPlex indexes:

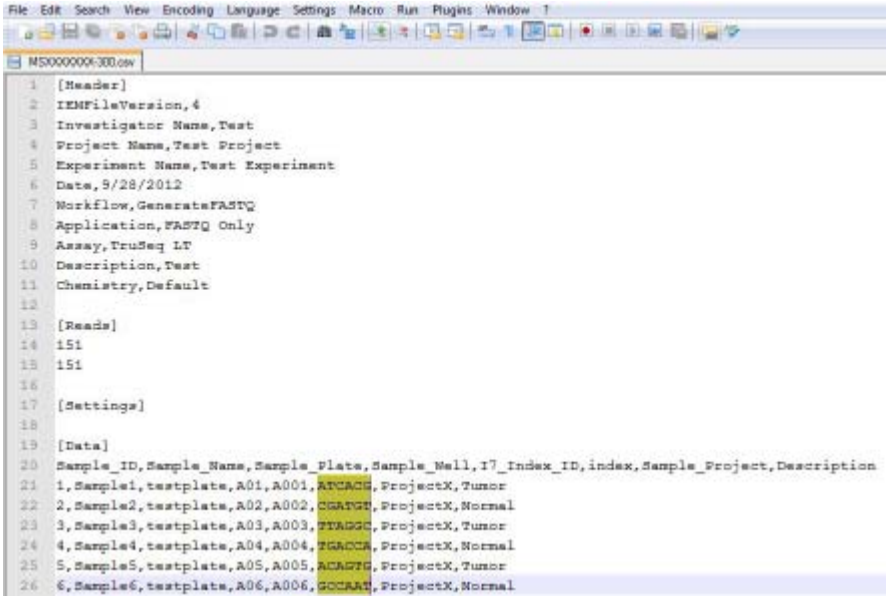
CAUTION

This guide includes information for kits containing two different sets of indexing primers. **Verify that you are referencing the information appropriate for your kit version before you proceed.**

Kits with indexing primers supplied in a blue plate include 8-bp indexes A01 through H12. See page 88 for index nucleotide sequence information.

Kits with indexing primers supplied in a clear plate include 8-bp indexes 1 through 96. See page 92 through page 97 for index nucleotide sequence information.

1 Open the Sample Sheet file in a text editor. For each sample, select the text for the 6-nucleotide index (highlighted below), and replace with the appropriate 8-nucleotide HaloPlex index sequence.



2 Save the edited Sample Sheet in an appropriate file location for use in the MiSeq platform run.

HiSeq platform sequencing run setup guidelines

Set up sequencing runs to perform an 8-nt index read using the *Cycles* settings shown in Table 32. Cycle number settings can be specified on the *Run Configuration* screen of the instrument control software interface after choosing *Custom* from the index type selection buttons.

Table 32 HiSeq platform Run Configuration screen Cycle Number settings*

Run Segment	Cycle Number
Read 1	100
Index 1 (i7)	9
Index 2 (i5)	0
Read 2	100

* Settings apply to v3.0 SBS chemistry.

Sequence analysis resources

Agilent's SureCall data analysis software is available to simplify the sequencing data analysis workflow after HaloPlex target enrichment. To learn more about this resource and download the SureCall software free of charge, visit www.agilent.com/genomics/surecall.

4 Appendix: Provisional Adaptor-Dimer Removal Protocol

Purify the enriched library pool using AMPure XP beads 80

This section contains a protocol for purification of the target-enriched library pool to remove adaptor-dimer molecules of approximately 125 bp size. Only do this protocol if electrophoretic analysis of the target-enriched library samples shows a peak at approximately 125 bp which represents a molar fraction of >10% of DNA in the sample (see page 70 to page 72.)

Purify the enriched library pool using AMPure XP beads

In this step, a 40-µL pool of target-enriched DNA libraries is purified using AMPure XP beads using manual sample processing.

NOTE This protocol requires a 0.2 mL tube-compatible magnetic separation device, such as the Agencourt SPRIPlate Super Magnet Plate, Agencourt p/n A32782, or equivalent.

1. Let the AMPure XP beads come to room temperature for at least 30 minutes.
2. Prepare 400 µL of 70% ethanol per sample, plus excess, for use in step 10.
3. Transfer 40 µL of each target-enriched library pool to a fresh 0.2-mL tube.
4. Mix the AMPure XP bead suspension well, until the suspension appears homogeneous and consistent in color.
5. For each sample to be purified, prepare a bead mix by combining 40 µL of nuclease-free water and 100 µL of the homogeneous AMPure XP bead suspension. Mix well, until the bead mix suspension appears homogeneous.
6. Add 140 µL of the homogeneous bead suspension prepared in step 5 to each 40-µL DNA sample. Vortex thoroughly.

   Using this bead-to-sample volume ratio is imperative to ensure optimal purification results.
7. Incubate samples for 5 minutes at room temperature with continuous shaking.

   Make sure the samples are properly mixing in the wells during the 5-minute incubation.
8. Spin briefly to collect the liquid, then place the tubes in the magnetic separation device. Wait for the solution to clear (approximately 5 minutes).
9. Keep the tubes in the magnetic device. Carefully remove and discard the cleared solution from each tube using a 200-µL pipette set to 180 µL. Do not touch the beads while removing the solution.
10. Continue to keep the tubes in the magnetic device while you add 200 µL of 70% ethanol into the tubes.

    Use fresh 70% ethanol for optimal results.

11 Wait for 30 seconds to allow any disturbed beads to settle, then remove the ethanol using a 200-µL pipette set to 200 µL.

12 Repeat step 10 and step 11 once for a total of two washes.

13 Remove any residual ethanol with a 20-µL volume pipette.

14 Air-dry the tubes with open lids at room temperature until the residual ethanol completely evaporates.

Make sure all ethanol has evaporated before continuing.

15 Remove tubes from the magnetic device and add 40 µL of 10 mM Tris-acetate or Tris-HCl buffer (pH 8.0) to each sample.

NOTE Use room-temperature Tris-acetate or Tris-HCl buffer for elution at this step.

16 Mix thoroughly by pipetting up and down 15 times using a 100-µL pipette set to 30 µL.

17 Incubate for 2 minutes at room temperature to allow elution of DNA.

18 Put the tube in the magnetic device and leave for 2 minutes or until the solution is clear.

19 Remove the cleared supernatant (approximately 40 µL) to a fresh tube. You can discard the beads at this time.

Stopping Point If you do not continue to the next step, samples may be stored at –20°C for long-term storage (up to one year). Avoid subjecting the stored DNA samples to multiple freeze-thaw cycles.

4 Appendix: Provisional Adaptor-Dimer Removal Protocol

Purify the enriched library pool using AMPure XP beads

5 Reference

Reference Information for Kits with Revised Index Configuration (indexing primers in blue plate) 84
Reference Information for Kits with Original Index Configuration (indexing primers in clear plate) 89
Qualitative analysis of enrichment by gel electrophoresis 98

This chapter contains reference information, including component kit contents, index sequences, and optional gel validation instructions.

CAUTION

This chapter contains two sets of index sequence and kit content information. **Verify that you are referencing the information appropriate for your kit version before you proceed.**

The first section covers kits with reconfigured indexing primers, typically received December, 2014 or later. The reconfigured primers A01–H12 are supplied in a blue plate. See page 84 through page 88 for details.

The second section covers kits with original indexing primer configuration, typically received before December, 2014. The original configuration includes primers 1–96, supplied in a clear plate. See page 89 through page 97 for details.

Reference Information for Kits with Revised Index Configuration (indexing primers in blue plate)

If your kit includes indexing primers in a clear plate, instead see page 89 for kit content and indexing primer information.

Kit Contents-Revised Configuration

HaloPlex Target Enrichment System Kits (revised index configuration) for custom designs include the component kits listed in Table 33 below. Kits for ClearSeq Disease Research Panels include the component kits listed in Table 34 below.

Table 33 HaloPlex Target Enrichment System Kit Contents-Custom Designs

Design Type	HaloPlex Target Enrichment System-ILM, Box 1*	HaloPlex Magnetic Beads Box 2
	Store at –20°C	Store at +4°C
HaloPlex 1-500 kb (up to 20,000 probes), ILM, 96 Reactions	5190-8050 **OR** 5190-8051†	5190-5386
HaloPlex 0.5-2.5 Mb OR <0.5 Mb with >20,000 probes, ILM, 96 Reactions	5190-8052 **OR** 5190-8053†	5190-5386
HaloPlex 2.6 Mb-5 Mb, ILM, 96 Reactions	5190-8054 **OR** 5190-8055†	5190-5386

* See Table 35 for list of included reagents.

† Part number 5190-8050, 5190-8052, or 5190-8054 is provided for the first order of a specific HaloPlex Probe design. Re-order kits, containing previously-purchased HaloPlex Probe designs, include Box 1 part number 5190-8051, 5190-8053, or 5190-8055.

Table 34 HaloPlex Target Enrichment System Kit Contents-ClearSeq Disease Research Panels

Design Type	HaloPlex Target Enrichment System-ILM, Box 1*	HaloPlex Magnetic Beads Box 2
	Store at –20°C	Store at +4°C
ClearSeq Cancer, ILM, 96 Reactions	5190-8057	5190-9224
ClearSeq Cardiomyopathy, ILM, 96 Reactions	5190-8059	5190-9224
ClearSeq AML, ILM, 96 Reactions	5190-8087	5190-9224
ClearSeq ICCG, ILM, 96 Reactions	5190-9164	5190-9224
ClearSeq Connective Disorder, ILM, 96 Reactions	5190-9168	5190-9224
ClearSeq Arrhythmia, ILM, 96 Reactions	5190-9162	5190-9224
ClearSeq Noonan Syndrome, ILM, 96 Reactions	5190-9170	5190-9224
ClearSeq Chromosome-X, ILM, 96 Reactions	5190-9166	5190-9224

* See Table 35 for list of included reagents.

The contents of the HaloPlex Target Enrichment System Box 1 (revised index configuration) included with each kit are detailed in the table below:

Table 35 HaloPlex Target Enrichment System Box 1 Contents

Included Reagents	Format
Hybridization Solution	bottle
Ligation Solution	bottle
Wash Solution	bottle
Capture Solution	bottle
SSC Buffer	bottle
RE Buffer	bottle
BSA Solution	tube with clear cap
DNA Ligase	tube with red cap
Enrichment Control DNA	tube with orange cap
Primer 1	tube with yellow cap
Primer 2	tube with blue cap
HaloPlex Indexing Primers	96-well plate with Indexing Primer A01 to H12 (blue plate)*
Enzyme Strip 1	8-well strip tube with green label
Enzyme Strip 2	8-well strip tube with red label
HaloPlex or ClearSeq Probe	tube with pink cap

* See Table 36 for a plate map.

Table 36 Plate map for HaloPlex Indexing Primers A01 through H12 provided in blue plate

	1	2	3	4	5	6	7	8	9	10	11	12
A	A01	A02	A03	A04	A05	A06	A07	A08	A09	A10	A11	A12
B	B01	B02	B03	B04	B05	B06	B07	B08	B09	B10	B11	B12
C	C01	C02	C03	C04	C05	C06	C07	C08	C09	C10	C11	C12
D	D01	D02	D03	D04	D05	D06	D07	D08	D09	D10	D11	D12
E	E01	E02	E03	E04	E05	E06	E07	E08	E09	E10	E11	E12
F	F01	F02	F03	F04	F05	F06	F07	F08	F09	F10	F11	F12
G	G01	G02	G03	G04	G05	G06	G07	G08	G09	G10	G11	G12
H	H01	H02	H03	H04	H05	H06	H07	H08	H09	H10	H11	H12

Nucleotide Sequences of HaloPlex Indexes (indexing primers in blue plate)

The nucleotide sequence of the 8-nucleotide index portion of each HaloPlex Indexing Primer (revised index configuration) is provided in the table below.

Table 37 HaloPlex Indexes, for indexing primers provided in blue 96-well plate

Index	Sequence	Index	Sequence	Index	Sequence	Index	Sequence
A01	ATGCCTAA	A04	AACTCACC	A07	ACGTATCA	A10	AATGTTGC
B01	GAATCTGA	B04	GCTAACGA	B07	GTCTGTCA	B10	TGAAGAGA
C01	AACGTGAT	C04	CAGATCTG	C07	CTAAGGTC	C10	AGATCGCA
D01	CACTTCGA	D04	ATCCTGTA	D07	CGACACAC	D10	AAGAGATC
E01	GCCAAGAC	E04	CTGTAGCC	E07	CCGTGAGA	E10	CAACCACA
F01	GACTAGTA	F04	GCTCGGTA	F07	GTGTTCTA	F10	TGGAACAA
G01	ATTGGCTC	G04	ACACGACC	G07	CAATGGAA	G10	CCTCTATC
H01	GATGAATC	H04	AGTCACTA	H07	AGCACCTC	H10	ACAGATTC
A02	AGCAGGAA	A05	AACGCTTA	A08	CAGCGTTA	A11	CCAGTTCA
B02	GAGCTGAA	B05	GGAGAACA	B08	TAGGATGA	B11	TGGCTTCA
C02	AAACATCG	C05	CATCAAGT	C08	AGTGGTCA	C11	CGACTGGA
D02	GAGTTAGC	D05	AAGGTACA	D08	ACAGCAGA	D11	CAAGACTA
E02	CGAACTTA	E05	CGCTGATC	E08	CATACCAA	E11	CCTCCTGA
F02	GATAGACA	F05	GGTGCGAA	F08	TATCAGCA	F11	TGGTGGTA
G02	AAGGACAC	G05	CCTAATCC	G08	ATAGCGAC	G11	AACAACCA
H02	GACAGTGC	H05	CTGAGCCA	H08	ACGCTCGA	H11	AATCCGTC
A03	ATCATTCC	A06	AGCCATGC	A09	CTCAATGA	A12	CAAGGAGC
B03	GCCACATA	B06	GTACGCAA	B09	TCCGTCTA	B12	TTCACGCA
C03	ACCACTGT	C06	AGTACAAG	C09	AGGCTAAC	C12	CACCTTAC
D03	CTGGCATA	D06	ACATTGGC	D09	CCATCCTC	D12	AAGACGGA
E03	ACCTCCAA	E06	ATTGAGGA	E09	AGATGTAC	E12	ACACAGAA
F03	GCGAGTAA	F06	GTCGTAGA	F09	TCTTCACA	F12	GAACAGGC
G03	ACTATGCA	G06	AGAGTCAA	G09	CCGAAGTA	G12	AACCGAGA
H03	CGGATTGC	H06	CCGACAAC	H09	CGCATACA	H12	ACAAGCTA

Reference Information for Kits with Original Index Configuration (indexing primers in clear plate)

If your kit includes indexing primers in a blue plate, instead see page 84 for kit content and indexing primer information.

Kit Contents-Original Configuration

The HaloPlex Target Enrichment System (original index configuration) includes the following component kits:

Table 38 HaloPlex Kit Contents

Design Type	HaloPlex Target Enrichment System-ILM, Box 1*	HaloPlex Magnetic Beads Box 2
	Store at –20°C	Store at +4°C
Custom 1-500 kb (up to 20,000 probes), ILMFST, 96 Reactions	5190-5385 **OR** 5190-5436†	5190-5386
Custom 0.5-2.5 Mb OR <0.5 Mb with >20,000 probes, ILM, 96 Reactions	5190-5534 **OR** 5190-5538†	5190-5386
Custom 2.6 Mb-5 Mb, ILM, 96 Reactions	5190-5536 **OR** 5190-5540†	5190-5386
Cancer Research, ILM, 96 Reactions	5190-6236	5190-5386
Cardiomyopathy Research, ILM, 96 Reactions	5190-6529	5190-5386
ClearSeq AML, ILM, 96 Reactions	5190-7735	5190-5386

* See Table 39 for list of included reagents.

† Part number 5190-5385, 5190-5534, or 5190-5536 is provided for the first order of a specific HaloPlex Probe design. Re-order kits, containing previously-purchased HaloPlex Probe designs, include Box 1 part number 5190-5436, 5190-5538, or 5190-5540.

The contents of the HaloPlex Target Enrichment System Box 1 (original index configuration) included with each kit are detailed in the table below:

Table 39 HaloPlex Target Enrichment System Box 1 Contents

Included Reagents	Format
Hybridization Solution	bottle
Ligation Solution	bottle
Wash Solution	bottle
Capture Solution	bottle
SSC Buffer	bottle
RE Buffer	bottle
BSA Solution	tube with clear cap
DNA Ligase	tube with red cap
Enrichment Control DNA	tube with orange cap
Primer 1	tube with yellow cap
Primer 2	tube with blue cap
HaloPlex Indexing Primer Cassettes	96-well plate with Indexing Primer Cassettes 1-96 (clear plate)*
Enzyme Strip 1	8-well strip tube with green label
Enzyme Strip 2	8-well strip tube with red label
HaloPlex or ClearSeq Probe	tube with pink cap

* See Table 40 for a plate map.

Table 40 Plate map for HaloPlex Indexing Primer Cassettes 1-96 provided in clear plate

	1	2	3	4	5	6	7	8	9	10	11	12
A	Index 1	Index 9	Index 17	Index 25	Index 33	Index 41	Index 49	Index 57	Index 65	Index 73	Index 81	Index 89
B	Index 2	Index 10	Index 18	Index 26	Index 34	Index 42	Index 50	Index 58	Index 66	Index 74	Index 82	Index 90
C	Index 3	Index 11	Index 19	Index 27	Index 35	Index 43	Index 51	Index 59	Index 67	Index 75	Index 83	Index 91
D	Index 4	Index 12	Index 20	Index 28	Index 36	Index 44	Index 52	Index 60	Index 68	Index 76	Index 84	Index 92
E	Index 5	Index 13	Index 21	Index 29	Index 37	Index 45	Index 53	Index 61	Index 69	Index 77	Index 85	Index 93
F	Index 6	Index 14	Index 22	Index 30	Index 38	Index 46	Index 54	Index 62	Index 70	Index 78	Index 86	Index 94
G	Index 7	Index 15	Index 23	Index 31	Index 39	Index 47	Index 55	Index 63	Index 71	Index 79	Index 87	Index 95
H	Index 8	Index 16	Index 24	Index 32	Index 40	Index 48	Index 56	Index 64	Index 72	Index 80	Index 88	Index 96

Nucleotide Sequences of HaloPlex Indexes (indexing primers in clear plate)

The nucleotide sequence of the 8-nucleotide index portion of each HaloPlex Indexing Primer Cassette (original index configuration) is provided in the Table 41 to Table 46 below.

Table 41 HaloPlex Indexes 1-16

Index Number	Sequence
1	AACGTGAT
2	AAACATCG
3	ATGCCTAA
4	AGTGGTCA
5	ACCACTGT
6	ACATTGGC
7	CAGATCTG
8	CATCAAGT
9	CGCTGATC
10	ACAAGCTA
11	CTGTAGCC
12	AGTACAAG
13	AACAACCA
14	AACCGAGA
15	AACGCTTA
16	AAGACGGA

Table 42 HaloPlex Indexes 17-32

Index Number	Sequence
17	AAGGTACA
18	ACACAGAA
19	ACAGCAGA
20	ACCTCCAA
21	ACGCTCGA
22	ACGTATCA
23	ACTATGCA
24	AGAGTCAA
25	AGATCGCA
26	AGCAGGAA
27	AGTCACTA
28	ATCCTGTA
29	ATTGAGGA
30	CAACCACA
31	CAAGACTA
32	CAATGGAA

Table 43 HaloPlex Indexes 33-48

Index Number	Sequence
33	CACTTCGA
34	CAGCGTTA
35	CATACCAA
36	CCAGTTCA
37	CCGAAGTA
38	CCGTGAGA
39	CCTCCTGA
40	CGAACTTA
41	CGACTGGA
42	CGCATACA
43	CTCAATGA
44	CTGAGCCA
45	CTGGCATA
46	GAATCTGA
47	GACTAGTA
48	GAGCTGAA

Table 44 HaloPlex Indexes 49-64

Index Number	Sequence
49	GATAGACA
50	GCCACATA
51	GCGAGTAA
52	GCTAACGA
53	GCTCGGTA
54	GGAGAACA
55	GGTGCGAA
56	GTACGCAA
57	GTCGTAGA
58	GTCTGTCA
59	GTGTTCTA
60	TAGGATGA
61	TATCAGCA
62	TCCGTCTA
63	TCTTCACA
64	TGAAGAGA

Table 45 HaloPlex Indexes 65-80

Index Number	Sequence
65	TGGAACAA
66	TGGCTTCA
67	TGGTGGTA
68	TTCACGCA
69	AACTCACC
70	AAGAGATC
71	AAGGACAC
72	AATCCGTC
73	AATGTTGC
74	ACACGACC
75	ACAGATTC
76	AGATGTAC
77	AGCACCTC
78	AGCCATGC
79	AGGCTAAC
80	ATAGCGAC

Table 46 HaloPlex Indexes 81-96

Index Number	Sequence
81	ATCATTCC
82	ATTGGCTC
83	CAAGGAGC
84	CACCTTAC
85	CCATCCTC
86	CCGACAAC
87	CCTAATCC
88	CCTCTATC
89	CGACACAC
90	CGGATTGC
91	CTAAGGTC
92	GAACAGGC
93	GACAGTGC
94	GAGTTAGC
95	GATGAATC
96	GCCAAGAC

Qualitative analysis of enrichment by gel electrophoresis

Enrichment products may be qualitatively analyzed by gel electrophoresis. Analyze 5 µL of each enriched library sample (enriched ECD sample or experimental enriched libraries) by electrophoresis on a Novex 6% polyacrylamide TBE pre-cast gel. See page 43 for additional gel analysis protocol recommendations.

Successful enrichment is indicated by the presence of a smear of amplicons from approximately 225 to 525 bp in each enrichment library lane. For some probe designs, low molecular weight (<150 bp) bands may also be visible, but should not be included in enriched sample quantitation. See Figure 10 for a sample gel analysis image.

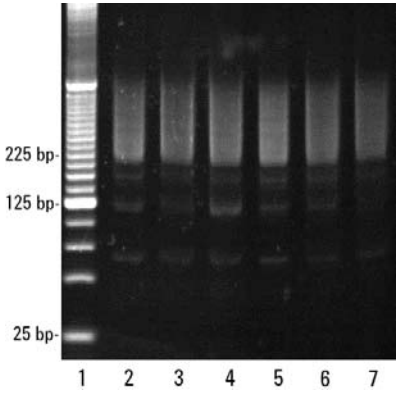


Figure 10 Validation of HaloPlex enrichment process by gel electrophoresis. Lane 1: 25-bp DNA ladder, Lanes 2-7: enriched library samples.

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In This Book

This guide contains information to run the HaloPlex Target Enrichment System automation protocol.

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